

**Ecophysiological Responses  
Of White Birch Seedlings to Soil Temperature and Phosphorus Supply  
Under Current and Doubled Carbon Dioxide Concentration**

**Gabriel Danyagri**

**Faculty of Forestry and the Forest Environment  
Lakehead University  
Thunder Bay  
Ontario**

## **A CAUTION TO THE READER**

This thesis has been through a formal process of review and comment by several committee members. As well, an external examiner has reviewed it.

It is made available for loan by the faculty for the purpose of advancing the practice of professional and scientific forestry.

The reader should realize that any opinions expressed in this thesis are those of the student, and do not necessarily reflect the opinions of the supervisor, the Faculty or the University.

## GENERAL ABSTRACT

To investigate the responses of some physiological and morphological traits to different soil temperature and phosphorus supply under current and doubled carbon dioxide concentration ( $[\text{CO}_2]$ ), white birch (*Betula papyrifera* Mash) seedlings were exposed to three soil temperatures ( $T_{\text{soil}}$ ) (7, 17 and 27°C) and three levels of phosphorus (P) (241, 493 and 951 mg/L) under current and doubled carbon dioxide concentration ( $[\text{CO}_2]$ ) (360 and 720  $\mu\text{mol mol}^{-1}$ , respectively). Morphological and physiological traits were measured after 2 and 4 months from the start of the treatments. The  $\text{CO}_2$  elevation significantly increased net rate of photosynthesis ( $P_n$ ) but such an increase did not lead to a corresponding increase in seedling growth. This was probably due to the negative effect of  $\text{CO}_2$  elevation on specific leaf area (SLA) as a consequence of changes in leaf anatomy and/or the accumulation of carbohydrates as the seedlings grew. The rate of photosynthesis measured at a common ambient  $[\text{CO}_2]$  showed that the  $\text{CO}_2$  stimulation of photosynthesis greatly declined over time. The foliar nutrient concentration in the elevated  $[\text{CO}_2]$  was also lower than that under ambient  $[\text{CO}_2]$  but seedlings grown in the elevated  $[\text{CO}_2]$  showed high nutrient-use-efficiencies.

Seedling height growth generally increased with increasing  $T_{\text{soil}}$  but the difference between the intermediate and high  $T_{\text{soil}}$  did not significantly differ under the elevated  $[\text{CO}_2]$ . The  $\text{CO}_2$  elevation partially mitigated the negative effect of low  $T_{\text{soil}}$  on seedling growth. There was also a substantial increase in total biomass due to the  $\text{CO}_2$  elevation at the intermediate and high  $T_{\text{soil}}$  but the low  $T_{\text{soil}}$  appeared to suppress biomass production probably due to its effect on nutrient and water uptake.

Phosphorus supply generally had a significant effect on seedling growth and biomass production but the effect varied with  $T_{\text{soil}}$  and  $[\text{CO}_2]$ . Height growth was significantly higher at the high P supply although it did not differ from the intermediate P at the high  $T_{\text{soil}}$  and elevated  $[\text{CO}_2]$  after 2 months. However, after 4 months, there was an increasing trend in height as P supply increased in both  $\text{CO}_2$  treatments. Biomass production was also higher at the intermediate and high P than at the low P supply under the ambient  $[\text{CO}_2]$  but the biomass was not significantly different between the low and intermediate P supply under the elevated  $[\text{CO}_2]$ . In summary, the growth of white birch seedlings was more sensitive to low  $T_{\text{soil}}$  than physiological traits. There was significant photosynthetic down-regulation in response to  $\text{CO}_2$  elevation and the down-regulation reduced the positive effect of  $\text{CO}_2$  elevation on the photosynthesis.

**Keywords:** White birch, Photosynthesis, carboxylation rate, gas exchange, foliar nutrient concentration, growth, biomass allocation, photosynthesis, transpiration, water-use-efficiency, nitrogen-use-efficiency.

## TABLE OF CONTENTS

	<b>Page</b>
<b>GENERAL ABSTRACT</b>	v
<b>LIST OF TABLES</b>	viii
<b>LIST OF FIGURES</b>	ix
<b>LIST OF ABBREVIATIONS</b>	x
<b>ACKNOWLEDGEMENT</b>	xi
<b>CHAPTER 1: GENERAL INTRODUCTION</b>	1
<b>CHAPTER 2: PHYSIOLOGICAL RESPONSES OF WHITE BIRCH SEEDLINGS TO SOIL TEMPERATURE AND PHOSPHORUS SUPPLY UNDER CURRENT AND DOUBLED CARBON DIOXIDE CONCENTRATION</b>	<b>5</b>
<b>ABSTRACT</b>	5
<b>INTRODUCTION</b>	7
<b>MATERIALS AND METHODS</b>	10
<b>RESULTS</b>	12
<b>DISCUSSION</b>	29
<b>CHAPTER 3: MORPHOLOGICAL RESPONSES TO SOIL TEMPERATURE AND PHOSPHORUS SUPPLY UNDER CURRENT AND DOUBLED CARBON DIOXIDE CONCENTRATION</b>	<b>34</b>
<b>ABSTRACT</b>	34
<b>INTRODUCTION</b>	36
<b>MATERIALS AND METHODS</b>	39
<b>RESULTS</b>	41
<b>DISCUSSION</b>	53
<b>CHAPTER 4: GENERAL DISCUSSION AND CONCLUSIONS</b>	57
<b>REFERENCES</b>	61
<b>APPENDIX 1 (LINEAR MODEL)</b>	72

## LIST OF TABLES

Table	Page
2.1. ANOVA table for $P_n$ , $P_{n360}$ , $g_s$ and IWUE	14
2.2. ANOVA table for $g_m$ , $C_i/C_a$ and $R_d$	18
2.3 ANOVA table for $V_{cmax}$ , $J$ and TPU	23
2.4 ANOVA table for foliar nutrient concentration after 4 months.	26
3.1 ANOVA table for height, RCD, leaf size and SLA	42
3.2 ANOVA table for SDM and RDM.	47
3.3 ANOVA table for TDM, SMR and RMR	51

## LIST OF FIGURES

<b>Figure</b>	<b>Page</b>
2.1. $P_n$ , $P_{n360}$ , $g_s$ and IWUE (mean $\pm$ S.E.).	16
2.2. $g_m$ , $C_i/C_a$ and $R_d$ (mean + S.E.).	20
2.3. $V_{cmax}$ , $J$ and TPU (mean + S.E.).	24
2.4. Foliar nutrient concentration, PUE and NUE (mean + S.E.).	28
3.1. Height, RCD, leaf size and SLA (mean + S.E.).	45
3.2. SDM and RDM (mean + S.E.).	48
3.3. TDM, SMR and RMR (mean + S.E.).	52

## LIST OF ABBREVIATIONS

[CO <sub>2</sub> ]	Carbon dioxide concentration
CO <sub>2</sub>	Carbon dioxide
$P_{n360}$	Photosynthesis at a common [CO <sub>2</sub> ]
$g_s$	Stomatal conductance
IWUE	Instantaneous water-use-efficiency
$J$	Apparent electron transport
$V_{cmax}$	Maximum carboxylation rate
TPU	Triose-phosphate utilization
$T_{soil}$	Soil temperature
SLA	Specific leaf area
P	Phosphorus
$g_m$	Mesophyll conductance
$C_i/C_a$	Intercellular and external CO <sub>2</sub> ratio
SDM	Shoot dry mass
RDM	Root dry mass
TDM	Total dry mass
SMR	Shoot mass ratio
RMR	Root mass ratio
PUE	Phosphorus use-efficiency
NUE	Nitrogen use-efficiency
N	Nitrogen
K	Potassium
SE	Standard error
$K_m$	Mass-based foliar potassium concentration
$K_a$	Area-based foliar potassium concentration
$P_m$	Mass-based foliar phosphorus concentration
$P_a$	Area-based foliar phosphorus concentration
$N_m$	Mass-based foliar nitrogen concentration
$N_a$	Area-based foliar nitrogen concentration
RCD	Root collar diameter
$P_n$	Net rate of photosynthesis



## ACKNOWLEDGMENTS

I express my heartfelt gratitude to my supervisor, Dr. Qing-Lai Dang, for his guidance, patience and constructive criticisms throughout the research. I would also like to acknowledge the support of my Committee members, Dr. Jian R. Wang and Dr. Chander Shahi. Their suggestions and constructive criticisms greatly contributed to the successful completion of this work. I also appreciate the technical support of Joan Lee, the greenhouse manager, during the course of the experiment. I am also grateful to my friends for their endless support and motivation during the course of my Master's study. Special thanks go my family for their assistance and encouragement which brought me this far.

# CHAPTER ONE

## GENERAL INTRODUCTION

The increases in atmospheric carbon dioxide concentration as a consequence of increased emissions from human activities (IPCC 2007) can have profound effects on photosynthesis and dry mass production of plants (Drake *et al.* 1997, Ward and Strain 1999, Zhang and Dang 2006). The primary productivity of all green organisms and ecosystems, particularly forest ecosystems, will likely increase due to enhanced photosynthesis and suppressed photorespiration under elevated CO<sub>2</sub> environments (Gifford 1982; Bazzaz 1990; Lawlor and Mitchell 2000). The increased photosynthetic rate will result in higher growth and biomass production in plants (Gifford 1982; Bazzaz 1990; Reddy *et al.* 2000). However, it has been generally observed that enhancement of photosynthetic rates in response to CO<sub>2</sub> elevation decline with time due to limitations to growth by other environmental factors, such as nutrient and soil temperature (Sage 1994; Poorter 1998; Oren *et al.* 2001; Poorter and Pérez-Soba 2001; Rogers and Ellsworth 2002; Zhang and Dang 2005), and soil moisture (Ambebe and Dang 2009).

Photosynthesis is an important determinant of plant growth rate through its influences on available photosynthates and the efficiency and extent at which the photosynthates are used productively by the plants (Farrar and Williams 1991). However, both photosynthetic rates and growth are also influenced by soil temperature and nutrient availability. Soil temperature affects root growth (Pastor *et al.* 1987; DeLucia *et al.* 1992; Paré *et al.* 1993; Folks *et al.* 1995; Peng and Dang 2003) thereby influencing the surface

area of roots for water and nutrient absorption and the leaf area for carbon assimilation (Aphalo *et al.* 2006). Soil temperature also affects root permeability and the water status of the plant which in turn affect stomatal conductance (Day *et al.* 1991, Zhang and Dang 2005; Lambers *et al.* 2008), therefore influencing the response of plants to CO<sub>2</sub> enrichment (Gavito *et al.* 2001). It has also been observed that low soil temperature inhibits the rate of shoot and leaf growth (Peng and Dang 2003). However, the response to low soil temperature differs among plant organs. For example, Lopushinsky and Kaufmann (1984) reported that low soil temperature reduced shoot growth but completely stopped root growth in Douglas-fir. The lack of information on the effect of T<sub>soil</sub> on white birch performance implies that further investigations into the physiological and morphological responses of plants to soil temperature and other environmental factors should allow more reliable predictions to be made of plant performance at sites with different soil temperatures.

Plant growth in the boreal forest is mostly limited by low nutrient availability which consequently limits the biomass production and carbon uptake in the ecosystem (Tamm 1991; Strömngren and Linder 2002). Nutrient availability affects the specific leaf area (SLA) and the total leaf area available for light interception and photosynthetic carbon assimilation (Lambers *et al.* 2008). The extent of plant growth stimulation by CO<sub>2</sub> elevation is reportedly lessened when plants are grown in nutrient poor environments (Ishizaki *et al.* 2003; Petterson *et al.* 1993). Past studies have generally focused on the effects of nitrogen availability and its effect on biomass production and carbon assimilation, especially with CO<sub>2</sub> enrichment or different temperatures (Ishizaki *et al.* 2003; Zhang and Dang 2006, Cao *et al.* 2007; Cao *et al.* 2008; Lou *et al.* 1994; Ambebe

*et al.* 2009). The present study focused on the interactive effects of soil temperature and P on physiological and morphological performance of white birch seedlings under ambient and elevated CO<sub>2</sub> concentration. P is one of the essential macronutrients required for the growth and development of higher plants (Lin *et al.* 2009). However, it is one of the limiting mineral nutrients in almost all soils due to its binding to soil mineral surfaces and fixation into organic forms (Kochian *et al.* 2004). P deficiency reduces leaf area development and shoot growth but increases the root/shoot ratio (Chaudhary *et al.* 2008; Whiteaker *et al.* 1976) as a result of reduction in leaf expansion and leaf initiation (Lynch *et al.* 1991; Nielsen *et al.* 2001). This indirectly reduces photosynthetic capacity and hydraulic conductance of the root system in plants (Chaudhary *et al.* 2008). P limitation also causes reduction in mesophyll capacity, stomatal conductance, photosynthetic quantum yield and rubisco activity or RuBP regeneration which reduces CO<sub>2</sub> assimilation (Rao and Terry 1989; Jacob and Lawlor 1991; Lin *et al.* 2009; Brooks 1986). The reduction in CO<sub>2</sub> assimilation rates in P deficient plants may be a direct result of the inhibition of triose-phosphate translocation across the chloroplast membrane at low P concentration in the stroma, the low demand for carbohydrates from sinks or a combination of both (Flügge *et al.* 1980; Sharkey 1985; Barrett and Gifford 1995).

With the increasing atmospheric CO<sub>2</sub> concentration and the subsequent rise in air temperature, changes in soil temperature may be inevitable which will likely affect nutrient availability and absorption, especially P. There is, however, a lack of information on the interactive effects of soil temperature and P supply on the physiological and morphological traits of white birch in ambient and elevated CO<sub>2</sub> concentration. Such information will improve our understanding on the responses of the boreal trees to the

changing climate associated with increasing atmospheric CO<sub>2</sub> concentration. It is hypothesized that the CO<sub>2</sub> elevation will enhance photosynthetic rate and total seedling biomass and the enhancement will be greater at the high soil temperature and high P supply. The objective of the study was to investigate the response of some physiological and morphological traits to different soil temperature and phosphorus supply under current and doubled [CO<sub>2</sub>].

## CHAPTER TWO

### PHYSIOLOGICAL RESPONSES OF WHITE BIRCH SEEDLINGS TO SOIL TEMPERATURE AND PHOSPHORUS SUPPLY UNDER CURRENT AND DOUBLED CARBON DIOXIDE CONCENTRATION

#### ABSTRACT

Increasing concentration of atmospheric CO<sub>2</sub> is predicted to impact both current and future ecosystems, especially boreal forest ecosystems. To investigate the physiological responses of white birch (*Betula papyrifera* Mash) seedlings to soil temperature (T<sub>soil</sub>) and phosphorus (P) supply under the current and elevated carbon dioxide concentration ([CO<sub>2</sub>]). Seedlings were grown at three T<sub>soil</sub> treatment (7, 17 and 27°C), three levels of P supply (241, 493 and 951 mg/L) and two [CO<sub>2</sub>] (360 and 720 μmol mol<sup>-1</sup>). In situ gas exchange measurements were done after 2 and 4 months from the start of the experiment. There was no significant difference in net photosynthesis (P<sub>n</sub>) between the intermediate and high T<sub>soil</sub> in the elevated [CO<sub>2</sub>] but P<sub>n</sub> was significantly higher at the intermediate T<sub>soil</sub> than the other T<sub>soil</sub> under the ambient [CO<sub>2</sub>] after 2 months. P<sub>n</sub> of seedlings grown at the high T<sub>soil</sub> was substantially higher after 4 months but it was down regulated in response to CO<sub>2</sub> elevation. When measured at the growth [CO<sub>2</sub>], seedlings under elevated [CO<sub>2</sub>] had higher P<sub>n</sub>, and instantaneous water-use-efficiency (IWUE) but lower stomatal conductance (g<sub>s</sub>) and lower intercellular CO<sub>2</sub>/ atmospheric CO<sub>2</sub> ratio (C<sub>i</sub>/C<sub>a</sub>) especially after 4 months. The CO<sub>2</sub> elevation induced maximum carboxylation rate (V<sub>cmax</sub>) down-regulation at the low P and intermediate T<sub>soil</sub> but CO<sub>2</sub> elevation induced apparent electron transport (J) down-regulation at the low and intermediate P and intermediate T<sub>soil</sub> after 4 months. However, the CO<sub>2</sub> elevation had no significant effect on V<sub>cmax</sub> after 2 months.

There was a transient down-regulation of triose-phosphate utilization (TPU) in response to CO<sub>2</sub> elevation at the intermediate T<sub>soil</sub> in the first measurement but not at the low and high T<sub>soil</sub>. The CO<sub>2</sub> elevation generally decreased the foliar nutrient but nutrient-use-efficiencies (photosynthetic nitrogen-use-efficiency (PNUE) and photosynthetic phosphorus-use-efficiency (PPUE)). The photosynthetic nutrient use-efficiency is an important functional trait that characterized species in relation to their physiology.

## INTRODUCTION

Photosynthetic carbon fixation by trees has a critical contribution to the productivity of forest ecosystems. Detailed information on how increases in atmospheric carbon dioxide concentration ( $[CO_2]$ ) will influence photosynthesis is critical for understanding how climate change would affect the structure, functioning and productivity of forest ecosystems (Cao *et al.* 2007). Photosynthetic responses to elevated  $[CO_2]$  can vary with other physiological and environmental conditions. For example, photosynthetic down-regulation is greater when plants are nutrient-stressed (Rogers *et al.* 1998, Saxe *et al.* 1998; Davey *et al.* 1999; Liozon *et al.* 2000; Zhang and Dang 2005, 2006; Cao *et al.* 2007). Most studies have shown that nutrient limitation reduces the beneficial effects of  $CO_2$  elevation on photosynthesis and growth (Saxe *et al.* 1998; Zhang and Dang 2006). However, most of past studies on nutrient have focused on nitrogen because it is the nutrient required in the largest quantity and is generally limiting to carbon assimilation (Chapin *et al.* 1987; Vaitkus *et al.* 1993; Li *et al.* 2004; Zhang and Dang 2006; Cao *et al.* 2007; Crous *et al.* 2008). Phosphorus is a key element regulating the physiological and biochemical reactions of photosynthesis (Lambers *et al.* 2008). In the present study, I have investigated the interactive effects of P supply and  $T_{soil}$  under ambient and elevated  $[CO_2]$  on some physiological parameters of white birch seedlings.

At elevated atmospheric  $CO_2$  concentrations, the ribulose-1,5-bisphosphate (RuBP) regeneration in photosynthesis is often limited by the rate of triose-phosphate utilisation (TPU) over the short term due to limited inorganic phosphate, but this limitation is



commonly relaxed after the plant is acclimated to the higher CO<sub>2</sub> concentration (Sage 1994).

Phosphorus is an essential element in plants, required for vital structural and metabolic functions and its deficiency can lead to a breakdown of plant membranes and reduce energy transfer within the plant (Oosterhuis *et al.* 2007). Phosphorus limitation indirectly reduces photosynthesis through its effects on leaf area development, photosynthetic capacity and hydraulic conductance of the root system (Chaudhary *et al.* 2008). Phosphate deficiency also decreases CO<sub>2</sub> assimilation through reduction in ribulose-1, 5-bisphosphate carboxylase/oxygenase (Rubisco) activity and RuBP regeneration (Brooks 1986; Jacob and Lawlor 1992; Lin *et al.* 2009). Studies using isolated chloroplasts and other *in-vitro* systems showed that phosphorus is involved in the activation of Rubisco (Heldt *et al.* 1978), modulation of ribulose-5-phosphate kinase and fructose-1,6-bisphosphate phosphatase (Leegood *et al.* 1985), the transport of triose-phosphate (TP) across the chloroplast membrane by the Pi-translocator and the regulation of photophosphorylation (Flügge *et al.* 1989). When inorganic phosphate (Pi) was withheld from plants, there was a substantial non-stomatal inhibition of photosynthesis (Rao and Terry 1989). In sugar beet, low-P treatment appeared to influence photosynthesis through RuBP regeneration rather than Rubisco activity (Rao and Terry 1995), indicating that low-P may affect photosynthetic rate differently in different species.

T<sub>soil</sub> affects the absorption of mineral nutrients by roots (Pastor *et al.* 1987; DeLucia *et al.* 1992; Paré *et al.* 1993). It has also been observed that low T<sub>soil</sub> inhibits the rates of shoot and leaf growth (Peng and Dang 2003) and root growth (Folks *et al.* 1995, Peng

and Dang 2003). Low  $T_{\text{soil}}$  can decrease both root growth and the formation of mycorrhizas (Domisch *et al.* 2001), thereby reducing the effective area for water and nutrient absorption (Aphalo *et al.* 2006). Cold soils decrease root permeability and increase water viscosity, leading to a decline in leaf conductance to water vapour and  $\text{CO}_2$  ( $g_l$ ) (Day *et al.* 1991, Lambers *et al.* 2008). This reduction in  $g_l$ , which is often associated with a decrease in shoot and leaf water potential ( $\Psi$ ), presumably limits net photosynthesis ( $P_n$ ). However, Teskey *et al.* (1983) observed a reduction in  $g_l$  at low  $T_{\text{soil}}$  without a decrease in leaf  $P_n$ . In this case, the mechanisms responsible for reduced  $g_l$  may be associated with: (a) hydraulic signals such as subtle changes in xylem flux (Teskey *et al.* 1983) that may reduce turgor of leaf epidermal cells but go undetected at the bulk leaf or shoot level; or (b) nonhydraulic signals between the roots and shoots involving hormones (Blackman and Davies 1985). Low  $T_{\text{soil}}$  can also decrease photosynthesis by reducing stomatal conductance ( $g_s$ ) to  $\text{CO}_2$  (Zhang and Dang 2005) and influence the acclimation of plants to elevated  $\text{CO}_2$  (Gavito *et al.* 2001).

$T_{\text{soil}}$  varies greatly in the boreal forest, ranging from near zero to over permafrost to  $35\text{ }^\circ\text{C}$  on south-facing slopes and newly burned areas (Bonan and Shugart 1989).  $T_{\text{soil}}$  can have substantial effects on the availability and absorption of P, particularly in the boreal forest where P is primarily absorbed through mycorrhizal association because it is immobile (Lambers *et al.* 2008). Declines in nutrient uptake at low soil temperatures cause a reduction in the size of the photosynthetic machinery (Aphalo *et al.* 2006). As the global climate changes in response to increasing atmospheric  $[\text{CO}_2]$ , changes in  $T_{\text{soil}}$  will be inevitable because of changes in snow cover and depth, air temperature and the duration of soil freezing (Aphalo *et al.* 2006). A small change in  $T_{\text{soil}}$  can have a profound

impact on the physiological processes of boreal plants (Cai and Dang 2002), including nutrient uptake and photosynthetic acclimation (Gavito *et al.* 2001; Camm and Harper 1991; Dang and Cheng 2004). A better understanding of interactions among [CO<sub>2</sub>], phosphorus supply and soil temperature on physiological traits of trees will provide insights for understanding the potential responses of boreal forests to rising atmospheric [CO<sub>2</sub>] and associated effects. The objective of the study was to investigate the interactive effects of [CO<sub>2</sub>], phosphorus supply and soil temperature on the photosynthetic functions of white birch (*Betula papyrifera* Marsh). It was hypothesized that the degree of photosynthetic down-regulation in response to CO<sub>2</sub> elevation would be greater under low T<sub>soil</sub> and low P supply.

## MATERIALS AND METHODS

### Plant materials

White birch seeds were germinated in the Lakehead University greenhouse (Thunder Bay, Ontario, Canada). The seeds were sown in horticultural trays filled with a mixture of peat moss and vermiculite (2:1 by volume). Seedlings of uniform height were transplanted to PVC containers (31.5 cm deep, 11 cm top diameter and 9.5 cm bottom diameter) after 4 weeks of germination and moved to treatment greenhouses as described below.

## **Experimental design**

The experiment comprised of two CO<sub>2</sub> concentrations, ambient (360 μmol mol<sup>-1</sup>) and elevated (720 μmol mol<sup>-1</sup>), three levels of P supply (241, 493 and 951 mg/L) and three T<sub>soil</sub> (7, 17 and 27° C). Nitrogen and potassium concentrations were kept at 221 and 150 mg/L, respectively, in all treatments. This was a split-split-split design. The CO<sub>2</sub> was the main plot, T<sub>soil</sub> the sub-plot and P the sub-sub-plot. The seedlings were fertilized twice a week. T<sub>soil</sub> was regulated by circulating temperature-controlled water between the containers within T<sub>soil</sub> control boxes. The boxes were insulated so that T<sub>soil</sub> was independent of the air temperature in the greenhouse. A detailed description of the system is given by Cheng et al. (2000). The day/night temperatures were 20 – 26/ 15 – 18°C and a day length of 16hr. The natural sunlight was supplemented using high-pressure sodium lamps on cloudy days, early mornings, and late evenings. Minimum illumination produced was about 660 μmol m<sup>-2</sup> s<sup>-1</sup>.

## **Gas exchange measurements**

Six seedlings per treatment combination were randomly selected for gas exchange measurements after two and four months of treatment. The measurements were taken with a CIRAS-1 open gas exchange system (PP-Systems, Hitchin Hertfordshire, U.K.) between 0900 and 1200hr on selected mature and nonshaded leaves. The environmental conditions in the leaf chamber (50% relative humidity (RH), 800 μ mol m<sup>-2</sup>s<sup>-1</sup> photosynthetically active radiation (PAR), and 26 °C air temperature) were automatically controlled by the system. The photosynthetic response curves to [CO<sub>2</sub>] were measured at 50, 150, 250, 300, 500, 700, 900 and 1500 μmol mol<sup>-1</sup> CO<sub>2</sub>.

## ***In vivo* biochemical activities of Rubisco**

*In vivo* maximal carboxylation rates ( $V_{\text{cmax}}$ ), photosynthetic electron transport rate ( $J$ ), triose-phosphate utilization (TPU), day-time dark respiration ( $R_d$ ) and mesophyll conductance ( $g_m$ ) were calculated from the  $A/C_i$  curve according to Farquhar (1980) and Harley and Sharkey (1991). The  $A/C_i$  curves were fit using the  $A/C_i$  curve fitting utility version 1.1 developed by Sharkey *et al.* (2007).

## **Leaf nutrient (N, P, K) assays**

The nutrient analysis was conducted at the Lakehead University Forest Soil Lab. Total nitrogen was analyzed using a LECO CNS 2000 principle. Total P and K were analyzed using nitric/hydrochloric acid digestion method (Goodfellow 2004). The mass-based nutrient concentrations were converted to area-based concentration by dividing the mass-based concentration by the specific leaf area. Photosynthetic Nitrogen- and P-use efficiencies (PNUE and PPUE, respectively) were calculated by dividing the  $P_n$  at the growth [ $\text{CO}_2$ ] by the area-based leaf N and P concentrations, respectively.

## **Data analysis**

The data were analysed using analysis of variance (ANOVA) with Data Desk 6.0 (Data Description, Ithaca, NY). When a factor or interactions between or among treatments for any parameter was significant, multiple comparisons of means were conducted using the Least Square Difference (LSD) method to identify treatment combinations that were significantly different from each other or one another.

## RESULTS

### Gas exchange

After two months of treatment, the interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  significantly ( $p < 0.05$ ) affected net photosynthesis ( $P_n$ ) measured at the corresponding growth  $[\text{CO}_2]$  (Table 2.1). The  $\text{CO}_2$  elevation increased  $P_n$  by 90.3% at the high  $T_{\text{soil}}$  and this  $\text{CO}_2$  stimulation completely offset the negative effect of the high  $T_{\text{soil}}$ . In contrast, the  $\text{CO}_2$  elevation had no significant effect on  $P_n$  at intermediate or low  $T_{\text{soil}}$  (Table 2.1, Fig 2.1A). After 4 months of treatment, the interaction of  $[\text{CO}_2]$  and  $T_{\text{soil}}$  became statistically insignificant ( $p > 0.10$ ) (Table 2.1). The  $\text{CO}_2$  elevation significantly ( $p < 0.05$ ) increased  $P_n$  at all  $T_{\text{soil}}$  (Fig. 2.1B).  $P_n$  increased with increases in  $T_{\text{soil}}$  under both  $[\text{CO}_2]$  (Fig. 2.1B).

**Table 2.1.** Probabilities from ANOVA for the effects of soil temperature ( $T_{\text{soil}}$ ) and phosphorus supply (P) under current and doubled  $[\text{CO}_2]$  on the rate of net photosynthesis at growth  $[\text{CO}_2]$  ( $P_n$ ), photosynthetic rate measured at a common  $[\text{CO}_2]$  ( $P_{n360}$ ), stomatal conductance to water ( $g_s$ ) and instantaneous water-use-efficiency (IWUE) in white birch seedlings. The seedlings were grown under two  $[\text{CO}_2]$  (360 and 720  $\mu\text{mol mol}^{-1}$ ), three  $T_{\text{soil}}$  (7, 17 and 27° C) and 3 levels of P supply (241, 493 and 951 mg/L). Measurements were taken 2 and 4 months after the start of the treatment.

Source of variation	$\text{CO}_2$	$T_{\text{soil}}$	$\text{CO}_2 \times T_{\text{soil}}$	P	$\text{CO}_2 \times \text{P}$	$T_{\text{soil}} \times \text{P}$	$\text{CO}_2 \times T_{\text{soil}} \times \text{P}$
After 2 months of treatment							
$P_n$	0.0140	<0.0001	0.0334	0.8698	0.8039	0.7916	0.4485
$P_{n360}$	0.0363	<0.0001	0.0411	0.9953	0.7894	0.8577	0.7935
$g_s$	0.6297	<0.0001	0.6069	0.3406	0.6523	0.924	0.9569
IWUE	0.0876	0.0282	0.8022	0.8757	0.7793	0.9901	0.9879
After 4 months of treatment							
$P_n$	0.0180	0.0078	0.9329	0.4150	0.8434	0.8486	0.6326
$P_{n360}$	<0.0001	0.0258	0.2246	0.4950	0.9930	0.9209	0.9887
$g_s$	0.0091	0.0103	0.0652	0.9889	0.8845	0.9918	0.9492
IWUE	0.0003	<0.0001	0.8969	0.8347	0.9889	0.7001	0.9801

After two months of treatment, the photosynthetic rate measured at a common, ambient  $[\text{CO}_2]$  ( $360 \mu\text{mol mol}^{-1}$ ) ( $P_{n360}$ ) was lower in the elevated  $[\text{CO}_2]$  than that under the ambient  $[\text{CO}_2]$  only under the intermediate  $T_{\text{soil}}$ , indicating that  $[\text{CO}_2]$  elevation resulted in photosynthetic down-regulation only at that  $T_{\text{soil}}$  (33%). Photosynthetic down-regulation after 4 months of treatment was statistically significant at all at 54.2, 50.8 and 60.2%, respectively at the low, intermediate and high  $T_{\text{soil}}$ . The low  $T_{\text{soil}}$  also resulted in a significant decline in  $P_{n360}$  (Fig. 2.1D).

Stomatal conductance ( $g_s$ ) increased with increase in  $T_{\text{soil}}$  but the  $[\text{CO}_2]$  had no significant effect on  $g_s$  after 2 months of treatment (Table 2.1, Fig. 2.1E). The interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  became marginally significant ( $p < 0.10$ ) after 4 months of treatment. The low  $T_{\text{soil}}$  suppressed  $g_s$  under ambient  $[\text{CO}_2]$  but not in the doubled  $[\text{CO}_2]$  treatment (Fig. 2.1F). However, the  $\text{CO}_2$  elevation significantly reduced  $g_s$  at the intermediate and high  $T_{\text{soil}}$  after the 4 months of treatment, but did not significantly affect  $g_s$  at the low  $T_{\text{soil}}$ .

The instantaneous water-use-efficiency (IWUE) was significantly influenced by the  $\text{CO}_2$  elevation and  $T_{\text{soil}}$  after 2 months of treatment (Table 2.1). The low  $T_{\text{soil}}$  significantly increased IWUE while the intermediate and high  $T_{\text{soil}}$  did not show any significant difference (Fig. 2.1G). After 4 months of treatment, IWUE decreased with increasing  $T_{\text{soil}}$  (Fig. 2.1H). The  $\text{CO}_2$  elevation also increased IWUE in the same pattern as observed after 2 months of treatment.



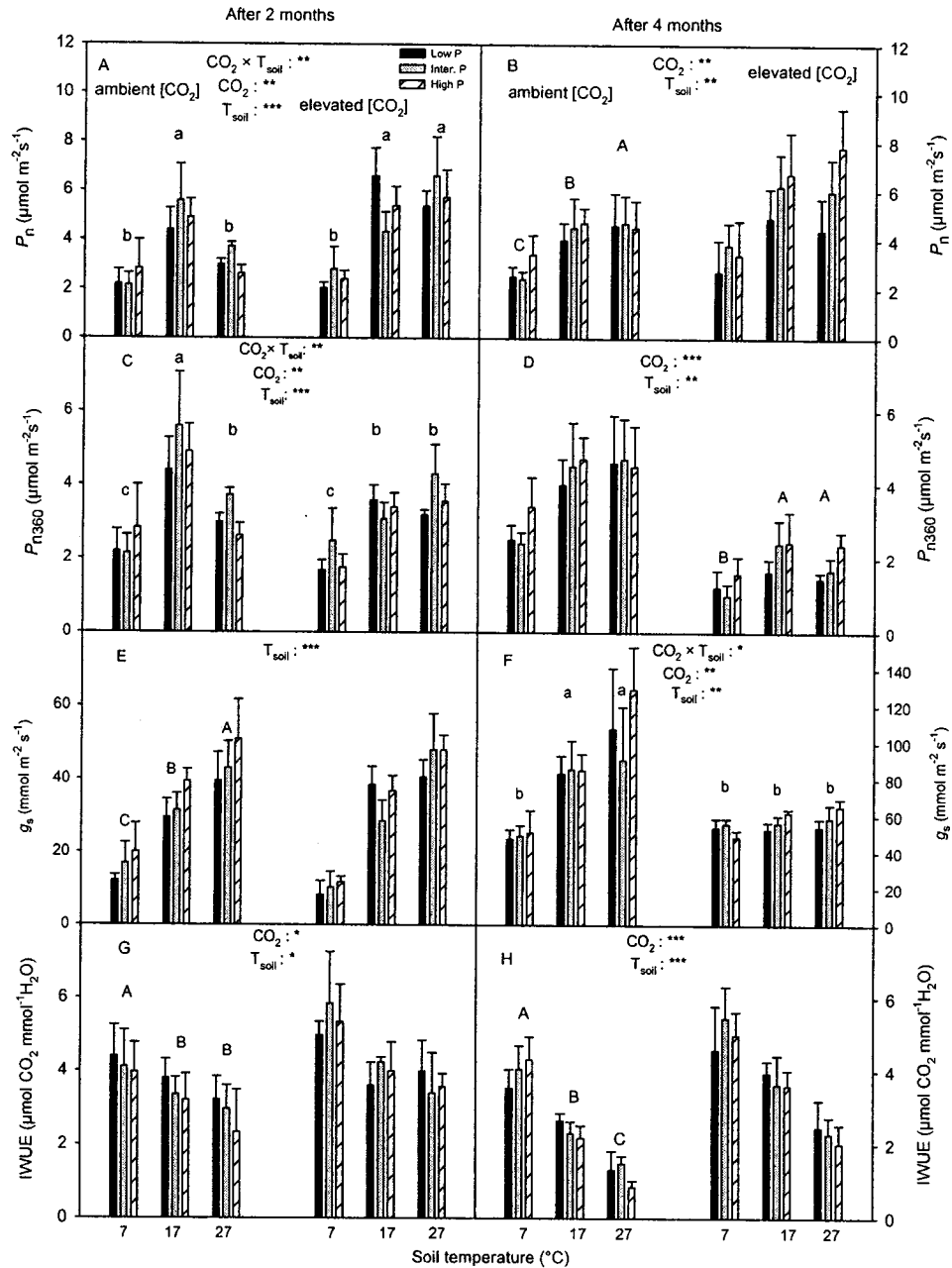


Figure 2.1. Effects of  $\text{CO}_2$  concentration ( $[\text{CO}_2]$ ), Soil temperature ( $T_{\text{soil}}$ ) and Phosphorus supply (P) on the rate of net photosynthesis ( $P_n$ ), photosynthetic rate measured at a common  $[\text{CO}_2]$  ( $P_{n360}$ ), stomatal conductance to water ( $g_s$ ) and instantaneous water-use-efficiency (IWUE) (mean + SE,  $n = 6$ ) in white birch seedlings after 2 and 4 months of treatment. The seedlings were grown under two  $[\text{CO}_2]$  (360 and 720  $\mu\text{mol mol}^{-1}$ ), three  $T_{\text{soil}}$  (7, 17 and 27° C) and 3 levels of P supply (241, 493 and 951 mg/L). The significance levels are: \*\*\* =  $P \leq 0.001$ , \*\* =  $P \leq 0.05$ , and \* =  $P \leq 0.10$ . Lower case letters above the bars represent interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  while upper case letters represent  $T_{\text{soil}}$  effect. Means with the same letter(s) are not significantly different from each other or one another.

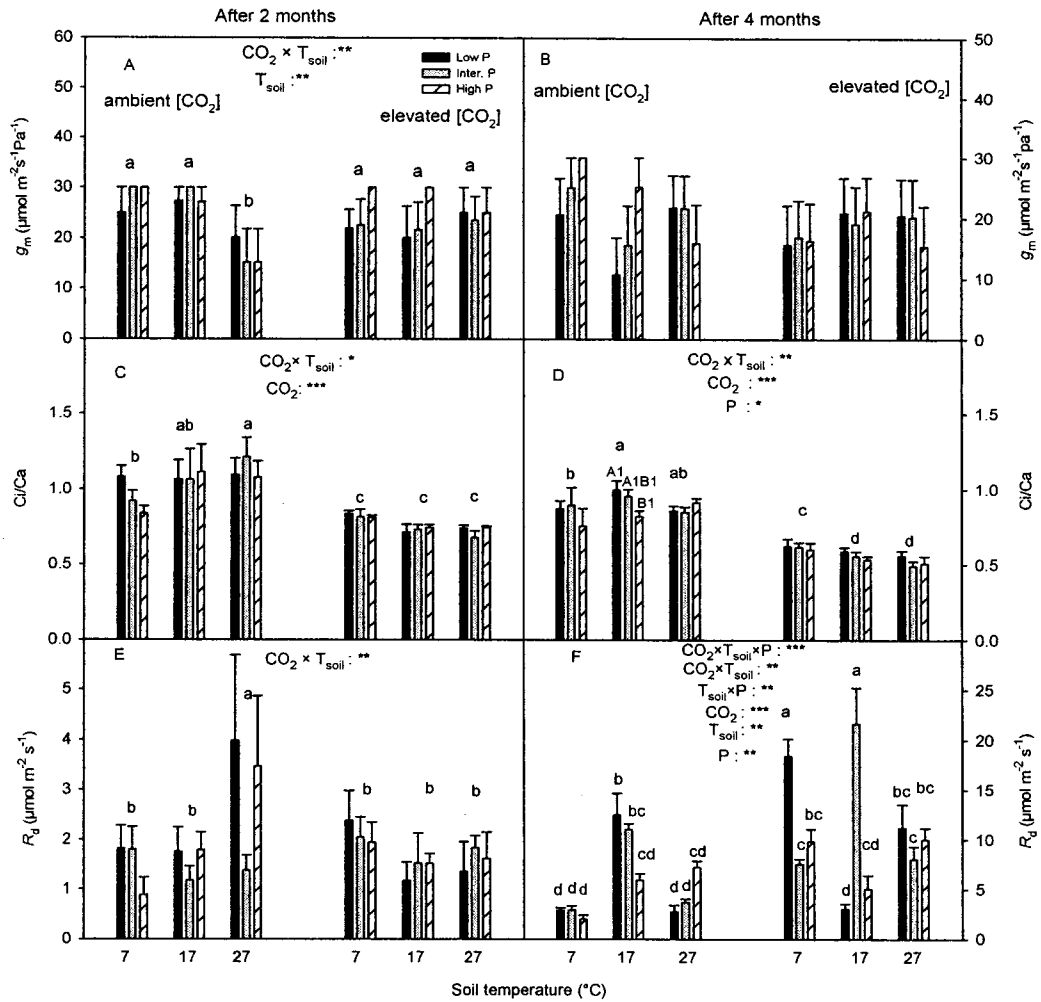
The  $[\text{CO}_2]$ - $T_{\text{soil}}$  interaction had a significant ( $p < 0.05$ ) effect on mesophyll conductance ( $g_m$ ) after 2 months of treatment (Table 2.2). The high  $T_{\text{soil}}$  significantly reduced  $g_m$  under the ambient but not under the elevated  $[\text{CO}_2]$ . The  $\text{CO}_2$  elevation, in contrast, increased  $g_m$  only at the high  $T_{\text{soil}}$  but did not significantly affect  $g_m$  at the low and intermediate  $T_{\text{soil}}$  (Fig. 2.2A). After 4 months of treatment however, none of the treatments significantly affected  $g_m$  (Table 2.2, Fig. 2.2B).

The intercellular/external  $[\text{CO}_2]$  ratio ( $C_i/C_a$ ) was significantly affected by the interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  after 2 months of treatment (Table 2.2).  $C_i/C_a$  were significantly higher the high than the low  $T_{\text{soil}}$  at the ambient  $[\text{CO}_2]$ , but  $T_{\text{soil}}$  had no significant effect under the doubled  $[\text{CO}_2]$  (Fig. 2.2C). The  $\text{CO}_2$  elevation significantly reduced  $C_i/C_a$  at all the  $T_{\text{soil}}$  (Fig. 2.2C). The  $[\text{CO}_2]$ -  $T_{\text{soil}}$  interaction remained significant after 4 months of treatment (Table 2.2). The  $C_i/C_a$  at the low  $T_{\text{soil}}$  was significantly lower than that at the intermediate and  $T_{\text{soil}}$  (Fig. 2.2D). The  $\text{CO}_2$  elevation significantly decreased  $C_i/C_a$  at all  $T_{\text{soil}}$  but the magnitude of decrease was higher at the intermediate and high  $T_{\text{soil}}$  (Fig. 2.2D).  $C_i/C_a$  was also significantly higher at the low than high P supply after the 4 months of treatment (Table 2.2, Fig. 2.2D).

**Table 2. 2.** Probabilities from ANOVA for the effects of  $T_{\text{soil}}$  and P supply under current and doubled  $[\text{CO}_2]$  on mesophyll conductance to  $\text{CO}_2$  ( $g_m$ ), intercellular  $\text{CO}_2$ /atmospheric  $\text{CO}_2$  ratio ( $C_i/C_a$ ) and daytime dark respiration rate ( $R_d$ ) in white birch seedlings. Other explanations are as in Table 1.

Source of variation	$\text{CO}_2$	$T_{\text{soil}}$	$\text{CO}_2 \times T_{\text{soil}}$	P	$\text{CO}_2 \times \text{P}$	$T_{\text{soil}} \times \text{P}$	$\text{CO}_2 \times T_{\text{soil}} \times \text{P}$
After 2 months of treatment							
$g_m$	0.9922	0.0416	0.0340	0.4663	0.3457	0.6412	0.9233
$C_i/C_a$	<b>&lt;0.0001</b>	0.8244	0.0955	0.8893	0.7684	0.8425	0.7762
$R_d$	0.3499	0.1531	0.0494	0.5276	0.2928	0.5915	0.4072
After 4 months of treatment							
$g_m$	0.4122	0.8217	0.1892	0.7825	0.5393	0.5213	0.925
$C_i/C_a$	<b>&lt;0.0001</b>	0.0258	0.2246	0.4950	0.9930	0.9209	0.9887
$R_d$	<b>0.0091</b>	0.0103	0.0652	0.9889	0.8845	0.9918	0.9492

The rate of daytime dark respiration ( $R_d$ ) was significantly ( $p < 0.05$ ) affected by the  $[\text{CO}_2]$ - $T_{\text{soil}}$  interaction after 2 months of treatment (Table 2.2).  $R_d$  was significantly higher at the high  $T_{\text{soil}}$  under the ambient  $[\text{CO}_2]$ , while there was no significant difference among other treatment (Fig. 2.2E). Although the high  $T_{\text{soil}}$  effect appeared to have primarily occurred under the ambient  $[\text{CO}_2]$  and the low and high P supply, the 3-way interaction was not significant ( $p > 0.10$ ). However, the interaction among  $[\text{CO}_2]$ ,  $T_{\text{soil}}$  and P supply became significant after 4 months of treatment (Table 2.2).  $R_d$  generally decreased with increasing P supply under the intermediate  $T_{\text{soil}}$  and ambient  $[\text{CO}_2]$ , but the difference between the low and intermediate, or between the intermediate and high P was not statistically significant (Fig. 2.2F). The  $\text{CO}_2$  elevation generally increased  $R_d$  at the low  $T_{\text{soil}}$ , but its effects under the other two  $T_{\text{soil}}$  varied with P supply (Fig. 2.2F).



**Figure 2.2.** Effects of CO<sub>2</sub> concentration ([CO<sub>2</sub>]), soil temperature (T<sub>soil</sub>) and phosphorus supply on mesophyll conductance to CO<sub>2</sub> ( $g_m$ ), intercellular CO<sub>2</sub>/atmospheric CO<sub>2</sub> ratio (Ci/Ca) and day-time dark respiration rate ( $R_d$ ) (mean +SE,  $n = 6$ ) in white birch seedlings. The lower case letters above the bars represent interaction among [CO<sub>2</sub>], T<sub>soil</sub> and P, or interaction between [CO<sub>2</sub>] and T<sub>soil</sub> and the upper case letter-number combination represent the effect of P. Other explanations are as in Figure 1.

## ***In vivo* biochemical and Rubisco activities**

After 2 months of treatments, both the low and high  $T_{\text{soil}}$  significantly reduced the maximum carboxylation rate ( $V_{\text{cmax}}$ ) of Rubisco but the degree of reduction was greater at the low  $T_{\text{soil}}$  (Table 2.3, Fig. 2.3A).  $[\text{CO}_2]$  and P supply did not significantly influence  $V_{\text{cmax}}$  after 2 months of treatment (Table 2.3). After 4 months of treatment, however, there was a significant ( $p < 0.001$ ) interaction among  $[\text{CO}_2]$ ,  $T_{\text{soil}}$  and P supply on  $V_{\text{cmax}}$  (Table 2.3). P supply had significant effects on  $V_{\text{cmax}}$  only under the intermediate  $T_{\text{soil}}$ , but the pattern of the response was opposite for the two  $[\text{CO}_2]$ . Under the ambient  $[\text{CO}_2]$ , the low P supply resulted in a significantly higher  $V_{\text{cmax}}$  and there was no significant difference between the intermediate and high P supply (Fig. 2.3B). Under the elevated  $[\text{CO}_2]$ , in contrast,  $V_{\text{cmax}}$  generally increased with increasing P supply although the difference between the low and intermediate P was not statistically significant (Fig. 2.3B). The  $\text{CO}_2$  elevation significantly increased  $V_{\text{cmax}}$  under the low  $T_{\text{soil}}$  and low P and the intermediate  $T_{\text{soil}}$  and high P but, significantly reduced  $V_{\text{cmax}}$  at the intermediate  $T_{\text{soil}}$  and low P (Fig. 2.3B). The intermediate  $T_{\text{soil}}$  resulted in a significantly higher  $V_{\text{cmax}}$  at the low P and ambient  $[\text{CO}_2]$ , but at the high P under the elevated  $[\text{CO}_2]$  (Fig. 2.3B).

The apparent electron transport rate ( $J$ ) measured after 2 months of treatment was significantly ( $p < 0.05$ ) affected by the  $[\text{CO}_2]$ - $T_{\text{soil}}$  interaction (Table 2.3). Under the ambient  $[\text{CO}_2]$ , the intermediate  $T_{\text{soil}}$  resulted in a significantly higher  $J$  while the low  $T_{\text{soil}}$  resulted in a significantly lower  $J$  (Fig. 2.3C). In contrast,  $J$  generally increased with  $T_{\text{soil}}$  under the elevated  $[\text{CO}_2]$  but the difference in  $J$  was statistically not significant between the low and intermediate  $T_{\text{soil}}$  (Fig. 2.3C).  $J$  was significantly reduced by the  $\text{CO}_2$  elevation under the intermediate  $T_{\text{soil}}$  after 2 months treatment. The interaction

among [CO<sub>2</sub>], T<sub>soil</sub> and P had a significant ( $p < 0.001$ ) effect on  $J$  after 4 months of treatment (Table 2.3). At the intermediate T<sub>soil</sub>, the intermediate P supply only differed from the low and intermediate P supply at the high T<sub>soil</sub> and all P supplies at the low T<sub>soil</sub> under the ambient [CO<sub>2</sub>] (Fig. 2.3D). The CO<sub>2</sub> elevation significantly increased  $J$  at the low P and low T<sub>soil</sub>, at the high P and intermediate T<sub>soil</sub>, and at the intermediate P and high T<sub>soil</sub> (Fig. 2.3D). However, the CO<sub>2</sub> elevation significantly reduced  $J$  in the low and intermediate P at the intermediate T<sub>soil</sub>.

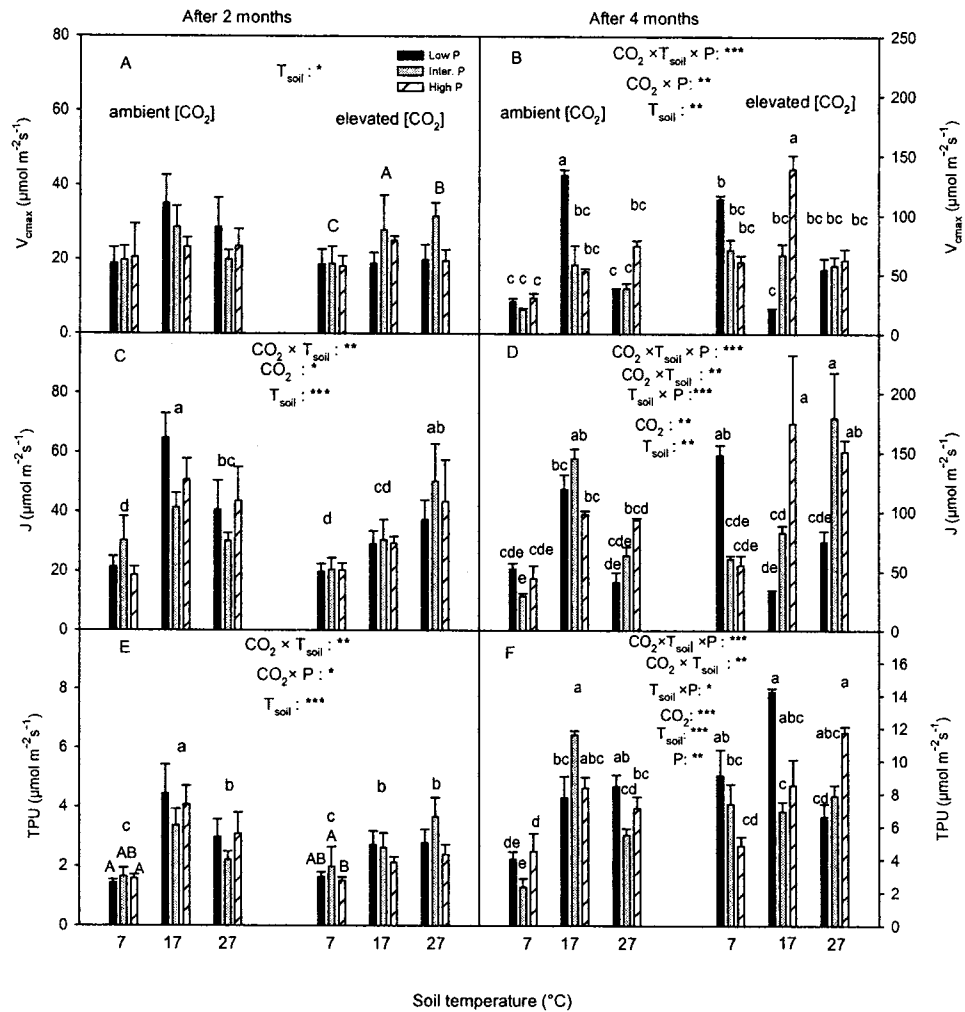
The [CO<sub>2</sub>]-T<sub>soil</sub> interaction significantly ( $p < 0.05$ ) influenced triose-phosphate utilization (TPU) after 2 months of treatment (Table 2.3). Under ambient [CO<sub>2</sub>], the TPU was highest at the intermediate T<sub>soil</sub> and lowest at the low T<sub>soil</sub> while TPU generally increased with T<sub>soil</sub> under the elevated [CO<sub>2</sub>] (Fig. 2.3E). The CO<sub>2</sub> elevation significantly reduced TPU at the intermediate T<sub>soil</sub>, but did not affect TPU at the low and high T<sub>soil</sub>. The interaction among [CO<sub>2</sub>], T<sub>soil</sub> and P supply after 4 months of treatment significantly ( $p < 0.001$ ) affected TPU (Table 2.3). The intermediate P supply had a significantly higher TPU at the intermediate T<sub>soil</sub> under the ambient [CO<sub>2</sub>], but the difference between the intermediate and high or between the high and low P supply was not statistically significant (Fig. 2.3F). Under the elevated [CO<sub>2</sub>], TPU increased at the low and intermediate P under the low T<sub>soil</sub>, at the low P and intermediate T<sub>soil</sub>, and at the high P and high T<sub>soil</sub> (Fig. 2.3F). The CO<sub>2</sub> elevation reduced TPU at the intermediate P and intermediate T<sub>soil</sub> and, at low P and high T<sub>soil</sub>, but significantly increased TPU at the low P and intermediate T<sub>soil</sub> (Fig. 2.3F).

**Table 2.3.** Probabilities from ANOVA for the effects of  $T_{\text{soil}}$  and P supply under current and doubled  $[\text{CO}_2]$  on the rate of maximum carboxylation ( $V_{\text{cmax}}$ ), rate of photosynthetic electron transport ( $J$ ) and triose-phosphate utilization (TPU) in white birch seedlings.

Other explanations are as in Table 1.

Source of variation	$\text{CO}_2$	$T_{\text{soil}}$	$\text{CO}_2 \times T_{\text{soil}}$	P	$\text{CO}_2 \times \text{P}$	$T_{\text{soil}} \times \text{P}$	$\text{CO}_2 \times T_{\text{soil}} \times \text{P}$
After 2 months of treatment							
$V_{\text{cmax}}$	0.4063	0.0653	0.7365	0.6942	0.1804	0.9787	0.4816
$J$	0.0803	<0.0001	0.0113	0.9317	0.3712	0.6272	0.5101
TPU	0.1286	<0.0001	0.0122	0.7939	0.1090	0.8773	0.8179
After 4 months of treatment							
$V_{\text{cmax}}$	0.1936	0.0024	0.2032	0.1160	0.0243	0.2908	<0.0001
$J$	0.0046	0.0022	0.0016	0.1502	0.3945	0.0005	0.0006
TPU	0.0012	<0.0001	0.0390	0.0250	0.4407	0.1057	<0.0001





**Figure 2.3.** Effects of  $\text{CO}_2$  concentration ( $[\text{CO}_2]$ ), soil temperature ( $T_{\text{soil}}$ ) and phosphorus supply on the rate of maximum carboxylation ( $V_{\max}$ ), rate of photosynthetic electron transport ( $J$ ) and triose-phosphate utilization (TPU) (mean + SE,  $n = 6$ ) in white birch seedlings. The lower case letters above the bars represent interactions among  $[\text{CO}_2]$ ,  $T_{\text{soil}}$  and P, or interactions between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  and the upper case letters represent interactions between  $[\text{CO}_2]$  and P or the effect of  $T_{\text{soil}}$ . Refer to Figure 1 for other explanations.

## **Foliar nutrient concentrations and nutrient use-efficiencies**

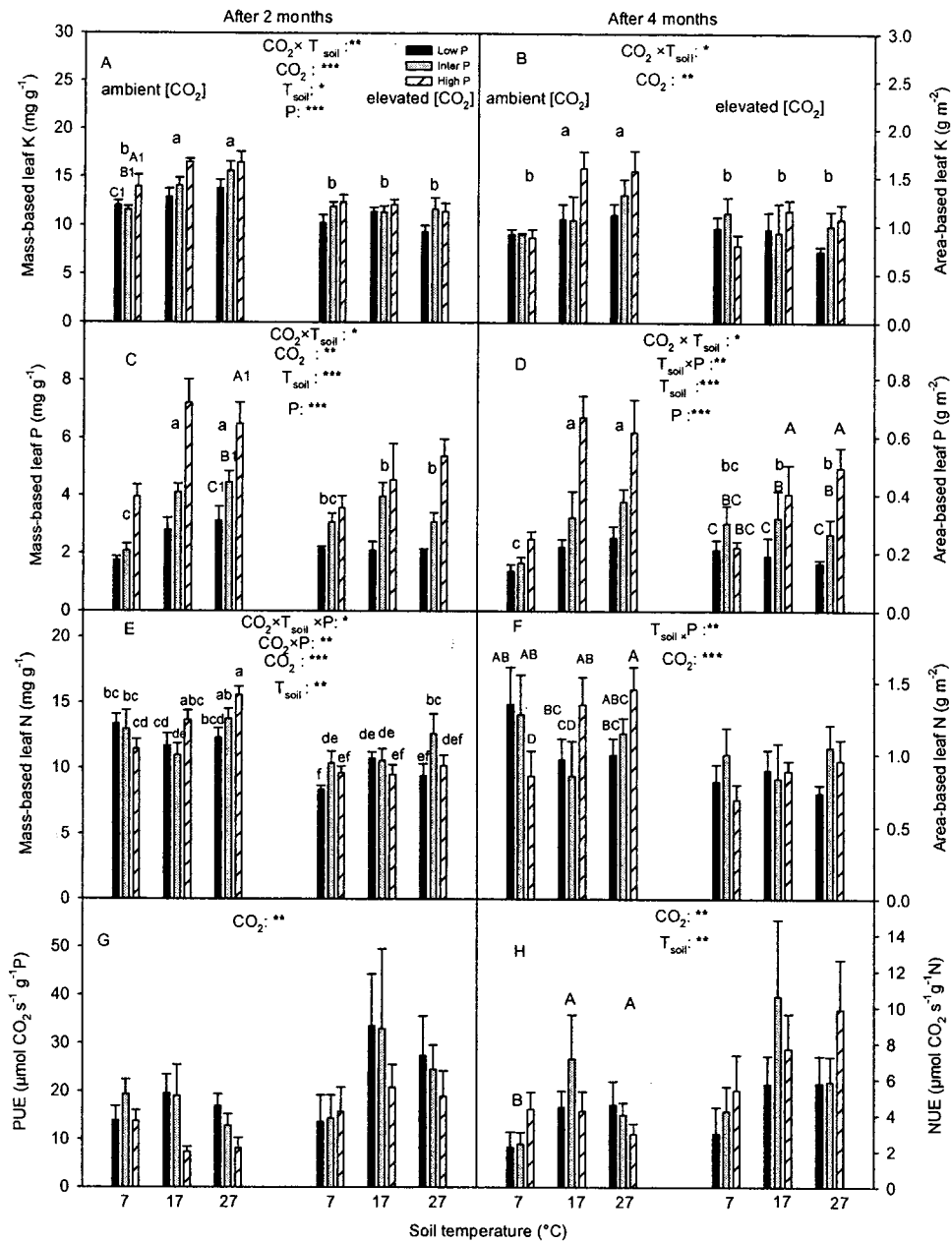
The  $[\text{CO}_2]$ - $T_{\text{soil}}$  interaction significantly influenced mass-based ( $K_m$ ) and area-based ( $K_a$ ) leaf potassium concentration after the 4 months of treatment (Table 2.4). The low  $T_{\text{soil}}$  significantly reduced both  $K_m$  and  $K_a$  while the  $\text{CO}_2$  elevation significantly reduced  $K_m$  and  $K_a$  at the intermediate and high  $T_{\text{soil}}$  (Figs. 2.4A and 2.4B). The  $K_m$  generally increased with increasing P supply but the  $K_a$  did not show significant response to P supply after the 4 months of treatment (Table 2.4, Fig. 2.4A).

The interaction between  $\text{CO}_2$  and  $T_{\text{soil}}$  significantly ( $p < 0.10$ ) affected both mass-based ( $P_m$ ) and area-based ( $P_a$ ) leaf phosphorus concentration (Table 2.4). Under ambient  $[\text{CO}_2]$ ,  $P_m$  and  $P_a$  were significantly lower at low than the intermediate and high  $T_{\text{soil}}$  while  $T_{\text{soil}}$  did not significantly affect  $P_m$  or  $P_a$  under elevated  $[\text{CO}_2]$  (Fig. 2.4C). The elevated  $\text{CO}_2$ , however, reduced  $P_m$  at both the intermediate and high  $T_{\text{soil}}$ . The  $P_m$  increased as the P supply increased at all  $T_{\text{soil}}$  (Table 2.4, Fig. 2.4C). However, the same trend was true for  $P_a$  only at the intermediate and high  $T_{\text{soil}}$  while P supply did not have a significant effect on  $P_a$  at the low  $T_{\text{soil}}$  (Fig. 2.4D).

**Table 2.4.** Probabilities from ANOVA for the effects of  $T_{\text{soil}}$  and P supply under current and doubled  $[\text{CO}_2]$  on mass-based leaf potassium concentration ( $K_m$ ), area-based leaf potassium concentration ( $K_a$ ), mass-based leaf phosphorus concentration ( $P_m$ ), area-based leaf phosphorus concentration ( $P_a$ ), mass-based leaf nitrogen concentration ( $N_m$ ), area-based leaf nitrogen concentration ( $N_a$ ), photosynthetic phosphorus use-efficiency (PPUE) and photosynthetic nitrogen use efficiency (PNUE) of white birch seedlings after 4 months of treatment. Other explanations are as in Table 2.1.

Source of variation	$\text{CO}_2$	$T_{\text{soil}}$	$\text{CO}_2 \times T_{\text{soil}}$	P	$\text{CO}_2 \times \text{P}$	$T_{\text{soil}} \times \text{P}$	$\text{CO}_2 \times T_{\text{soil}} \times \text{P}$
$K_m$	<0.0001	0.0967	0.0064	0.0002	0.2653	0.6605	0.6798
$K_a$	0.0329	0.1265	0.0956	0.1215	0.3877	0.1349	0.9885
$P_m$	0.0160	0.0003	0.0618	<0.0001	0.1649	0.3363	0.4456
$P_a$	0.1410	0.0012	0.1046	<0.0001	0.1378	0.0316	0.6929
$N_m$	<0.0001	0.0234	0.3268	0.1756	0.0548	0.1137	0.0609
$N_a$	0.0014	0.6367	0.7479	0.7249	0.4644	0.0271	0.5478
PUE	0.0218	0.2474	0.2109	0.1773	0.9743	0.7295	0.9954
NUE	0.0051	0.0146	0.5792	0.2863	0.4205	0.3170	0.7479

Mass-based leaf nitrogen concentration ( $N_m$ ) was significantly ( $p < 0.10$ ) affected by the interaction among  $[CO_2]$ ,  $T_{soil}$  and P supply (Table 2.4). Under ambient  $[CO_2]$ ,  $N_m$  was significantly higher at high P and high  $T_{soil}$  it was highest at high  $T_{soil}$  and intermediate P supply under the elevated  $[CO_2]$  (Fig. 2.4E). The  $CO_2$  elevation generally decreased  $N_m$  (Fig. 2.4E). The interaction between  $T_{soil}$  and P supply had a significant ( $p < 0.05$ ) effect on  $N_a$  after 4 months of treatment (Table 2.4).  $N_a$  was significantly lower at the high P at the low  $T_{soil}$ , while there was no significant difference between the low and intermediate P under the intermediate and high  $T_{soil}$  (Fig. 2.4E). The  $CO_2$  elevation significantly reduced  $N_a$ . Elevated  $[CO_2]$  significantly ( $p < 0.05$ ) increased both photosynthetic phosphorus use-efficiency (PPUE) and photosynthetic nitrogen use-efficiency (PNUE) (Table 2.4, Figs. 2.4G and 2.4H). However, low  $T_{soil}$  significantly reduced the PNUE (Fig 2.4H).



**Figure 2.4.** Effects of  $\text{CO}_2$  concentration ( $[\text{CO}_2]$ ), soil temperature ( $T_{\text{soil}}$ ) and phosphorus supply (P) on mass-based leaf potassium concentration ( $K_m$ ), area-based leaf potassium concentration ( $K_a$ ), mass-based leaf P concentration ( $P_m$ ), area-based leaf P concentration ( $P_a$ ), mass-based leaf nitrogen concentration ( $N_m$ ), area-based leaf nitrogen concentration ( $N_a$ ), photosynthetic phosphorus use-efficiency (PPUE) and photosynthetic nitrogen use efficiency (PNUE) (mean + SE,  $n = 6$ ) of birch seedlings after 4 months of treatment. The lower case letters above the bars represent interaction among  $[\text{CO}_2]$ ,  $T_{\text{soil}}$  and P, or interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$ ; the upper case letters above the bars represent interaction between  $T_{\text{soil}}$  and P, or the effect of  $T_{\text{soil}}$  and the upper case letter-number combination represent the effect of P. Other explanations are as in Figure 1.

## DISCUSSION

Our findings did not support the hypothesis that low P supply increases the degree of CO<sub>2</sub> elevation induced photosynthetic down-regulation. However, the CO<sub>2</sub> elevation did cause down-regulation of photosynthetic rate measured at a common ambient [CO<sub>2</sub>], but the degree of down-regulation did not vary with P supply. Arp (1991) suggested that photosynthetic down-regulation is a phenomenon associated with plants grown in pots where the root growth and photosynthate demand are restricted by the pots. However, photosynthetic down-regulation caused by elevated [CO<sub>2</sub>] have also been observed in some field studies (Tissue *et al.* 1999; Li *et al.* 2004; Rogers and Ellsworth 2002; Crous *et al.* 2008). Hence, photosynthetic down-regulation is not necessarily a characteristic response of plants grown in pots to CO<sub>2</sub> elevation as described in some literature (Long and Drake 1991; Long *et al.* 2004). Its occurrence may depend on other environmental conditions that the plants are experiencing. We observed that the magnitude of the photosynthetic down-regulation in response to CO<sub>2</sub> elevation was higher at the high T<sub>soil</sub> which supports our hypothesis that photosynthetic down-regulation will be higher at the high T<sub>soil</sub>. The insignificant photosynthetic down-regulation with P supply might have resulted from the constant amount of nitrogen supplied at all P levels. Studies found that photosynthetic rates of plants are more closely related to nitrogen because it is a major component of Rubisco and other photosynthetic enzymes and structures (Lewis 1994; Bond *et al.* 1999; Ripullone *et al.* 2003). However, phosphorus is a key element regulating the physiological and biochemical reactions of photosynthesis (Lambers *et al.* 2008) and it's essential for structural and metabolic functions (Oosterhuis *et al.* 2007).

We observed that the elevated [CO<sub>2</sub>] stimulation of  $P_n$  was higher at the high  $T_{soil}$  but the magnitude of CO<sub>2</sub> stimulation greatly declined with time. The measurement two months after the start of the treatment showed that CO<sub>2</sub> elevation increased  $P_n$  by 48% but the stimulation declined to 24% after 4 months of treatment. This is consistent with early observations that early enhancement of photosynthesis by elevated CO<sub>2</sub> may not be sustained over time (Sage 1994; Poorter 1998; Oren *et al.* 2001; Poorter and Pérez-Soba 2001; Rogers and Ellsworth 2002; Norby and Iversen 2006). The unsustainable photosynthetic enhancement by elevated [CO<sub>2</sub>] may be due to the influence of environmental factors such as nutrient limitation (Nowak *et al.* 2004; Warren and Dreyer 2006) and or unfavourable soil temperatures (Zhang and Dang 2005). Oren *et al.* (2001) and Lüo *et al.* (2004) reported that lack of photosynthetic enhancement by elevated [CO<sub>2</sub>] is especially apparent in low-nutrient ecosystems and is strongly related to the availability and root exploitation of limiting nutrients. We conclude that the decline of the photosynthetic enhancement after the 4 months of treatment might have resulted from the lower leaf nutrient concentration in the elevated [CO<sub>2</sub>] grown plants.

Elevated [CO<sub>2</sub>] suppresses  $g_s$  in plants (Bunce 1992; Rey and Jarvis 1998; Zhang and Dang 2005; Zhang and Dang 2006; Cao and Dang 2007; Crous *et al.* 2008; Lambers *et al.* 2008). Our initial observation of insignificant CO<sub>2</sub> effect is in contrast with this theory but supports the review of Saxe *et al.* (1998) who did not find significant decreases in  $g_s$  in response to CO<sub>2</sub> enrichment in trees, particular woody coniferous trees. Ellsworth *et al.* (1995) also found that  $g_s$  of *Pinus taeda* did not show reduced  $g_s$  after 80 days of exposure to elevated CO<sub>2</sub>, although plants in the elevated CO<sub>2</sub> did exhibit transient adjustment in  $g_s$  and reduction in water loss. Ellsworth (1999) reported that there are

exceptions to the general rule that  $g_s$  declines at elevated  $[\text{CO}_2]$  where the controlling mechanisms (guard cells) appear to be insensitive to elevated  $[\text{CO}_2]$ . The stomatal conductance however, was reduced by the  $\text{CO}_2$  elevation after 4 months of treatment which is in agreement with the theory that elevated  $\text{CO}_2$  decreases  $g_s$ .

We observed down-regulation of  $V_{\text{cmax}}$  at the low P supply and intermediate  $T_{\text{soil}}$  in response to the  $\text{CO}_2$  elevation. Decreased  $V_{\text{cmax}}$  could be associated with decrease in the activation state and amount of Rubisco (Sage *et al.* 1987) when the inorganic phosphate (Pi) for the synthesis of adenosine triose-phosphate (ATP) is locked in phosphorylated intermediates (Rogers *et al.* 1994). This reduces the rate of  $\text{CO}_2$  assimilation in P deficient plants (Brooks 1986; Jacob and Lawlor 1991; Lin *et al.* 2009) but our study did show significantly varied  $P_n$  in response to P levels, probably due to constant N supply.  $V_{\text{cmax}}$  is particularly important because a greater  $V_{\text{cmax}}$  increases the efficiency of net  $\text{CO}_2$  uptake by decreasing  $\text{CO}_2$  loss and diverting ATP and nicotinamide adenine dinucleotide (phosphate) (NADPH) away from photorespiratory metabolism to photosynthetic assimilation (Long 1991; Long and Drake 1991).

The insignificant  $\text{CO}_2$  elevation effect on  $V_{\text{cmax}}$  two months after the start of treatment is in agreement with the findings of Campbell *et al.* (1988) who reported similar results in soybean after a short-term exposure to  $\text{CO}_2$  enrichment. Other studies reported that  $V_{\text{cmax}}$  increased with increased  $\text{CO}_2$  concentration (Long 1991; Long and Drake 1992; Zhang and Dang 2006). Rey and Jarvis (1998) however, reported a significantly lower  $V_{\text{cmax}}$  in trees grown in elevated  $[\text{CO}_2]$  than those in ambient  $[\text{CO}_2]$  over a growing season.



Our results showed a transient down-regulation of triose-phosphate utilization (TPU) in response to the CO<sub>2</sub> elevation at the intermediate T<sub>soil</sub>. TPU is an indicator of sink strength. TPU was significantly lower at the elevated [CO<sub>2</sub>] than the ambient [CO<sub>2</sub>] at the intermediate T<sub>soil</sub> in the first measurement time but not at the low and high T<sub>soil</sub>. However, this effect became insignificant in the second measurement. The effect of CO<sub>2</sub> elevation on the rate of photosynthesis is often interpreted in terms of three general classes of biochemical limitations: Rubisco activity, RuBP regeneration capacity and the capacity for triose-phosphate utilization (Farquhar et al. 1980; Sharkey 1985). We observed a marginal up-regulation of apparent electron transport in response to CO<sub>2</sub> elevation at the high T<sub>soil</sub> but not at the low and intermediate T<sub>soil</sub>. This might have minimised the feedback inhibition on electron transport from TPU (Socias *et al.* 1993). Therefore, the lack of photosynthetic down-regulation at the high T<sub>soil</sub> after 2 months of treatment might have been the result of increased biochemical activities affecting CO<sub>2</sub> assimilation.

Our data show that the CO<sub>2</sub> elevation reduced foliar nutrient concentration but increased their use efficiency. The lower leaf nutrient concentration in the elevated [CO<sub>2</sub>] is consistent with the theory that elevated [CO<sub>2</sub>] can lead to the depletion of nutrient resources in plants unless it is replenished (Patterson and McDonald 1994). Zhang and Dang (2006) and Cao *et al.* (2007) also reported a lower leaf nutrient concentration in elevated [CO<sub>2</sub>]. Lower nutrient concentration in the elevated [CO<sub>2</sub>] might be due to dilution in whole plant nutrient content as the plant increased in size without a corresponding increase in nutrient concentration supplied. The nutrient-use-efficiencies of both P and N were generally higher under the elevated [CO<sub>2</sub>] than the ambient [CO<sub>2</sub>]

which is in agreement with earlier observation of increased NUE and PUE by elevated  $[\text{CO}_2]$  (Zhang and Dang 2006).

In conclusion, the rates of photosynthesis in white birch seedlings were stimulated by the  $\text{CO}_2$  elevation. However, down-regulation in the rates of photosynthesis in response to the elevated  $[\text{CO}_2]$  was evident and the magnitude of down-regulation was greater at the high  $T_{\text{soil}}$ . The biochemical results ( $V_{\text{cmax}}$ ,  $J$  and TPU) suggest that the biochemical activities of the seedlings are more sensitive to the P supply than the gas exchange processes. The P supply had significant effects on  $V_{\text{cmax}}$ ,  $J$  and TPU over time without a significant effect on the photosynthetic rates of the seedlings. However, we observed that  $P_n$  greatly down-regulated at high  $T_{\text{soil}}$  in response to  $\text{CO}_2$  elevation over time, which means that greater down-regulation of  $P_n$  in response to the increasing atmospheric  $[\text{CO}_2]$  might occur at high  $T_{\text{soil}}$ .

## CHAPTER THREE

# MORPHOLOGICAL RESPONSES OF WHITE BIRCH SEEDLINGS TO SOIL TEMPERATURE AND PHOSPHORUS SUPPLY UNDER CURRENT AND DOUBLED CARBON DIOXIDE CONCENTRATION

### ABSTRACT

Increases in carbon dioxide concentration ( $[CO_2]$ ) can increase plant growth but the stimulation of growth can be influenced by environmental factors such as nutrients and soil temperature ( $T_{soil}$ ). To better understand the performance of boreal trees under future atmospheric  $[CO_2]$ , white birch (*Betula papyrifera* Mash) seedlings were subjected to three  $T_{soil}$  (7, 17 and 27°C) and three phosphorus (P) supplies (241, 493 and 951 mg/L) under ambient and elevated  $[CO_2]$ . Seedling height, root collar diameter (RCD) and biomass were measured two and four months after the start of the experiment. The  $CO_2$  elevation stimulated height growth and partially mitigated the negative effect of low  $T_{soil}$  on height growth but the magnitude of the stimulation declined over time. There appeared to be a shift in the optimum  $T_{soil}$  (from high to low) for height growth with  $CO_2$  elevation. Height growth increased with increasing  $T_{soil}$  under the ambient  $[CO_2]$ . Under the elevated  $[CO_2]$  however, the height growth was not significantly different between the intermediate and high  $T_{soil}$ . Height growth at low and intermediate P supply was generally lower than the high P and did not appear to be significantly different from each other especially at the low and intermediate  $T_{soil}$  under the ambient  $[CO_2]$  and the low  $T_{soil}$  under the elevated  $[CO_2]$  after 2 months. The  $CO_2$  elevation also stimulated diameter growth and the magnitude of stimulation was greater at the intermediate  $T_{soil}$  than the other two  $T_{soils}$  after 2 months but the effect of  $T_{soil}$  disappeared after 4 months. After 2

months, the total biomass at all P supplies did not significantly differ in the ambient [CO<sub>2</sub>]. However, the intermediate and high P supply significantly increased total biomass after 4 months in the ambient [CO<sub>2</sub>]. The CO<sub>2</sub> elevation increased the shoot mass ratio (SMR) but did not affect root mass ratio (RMR) while the low T<sub>soil</sub> decreased SMR after 2 months.

## INTRODUCTION

The warming of the global climate is unequivocal as a consequence of the increased emissions of carbon dioxide through human activities since the pre-industrial time (IPCC 2007). Increases in atmospheric [CO<sub>2</sub>] can have profound effects on photosynthesis and dry mass production of plants (Drake *et al.* 1997; Ward and Strain 1999; Zhang and Dang 2006). The response, however, may be influenced by other environmental factors such as nutrient supply and soil temperatures (Nowak *et al.* 2004; Zhang and Dang 2005; Warren and Dreyer 2006). Many studies have investigated the effect of CO<sub>2</sub> elevation on the physiological responses of boreal trees (Warren and Adams 2001; Zhang and Dang 2005; Zhang and Dang 2006; Cao *et al.* 2007; Crous *et al.* 2008; Lin *et al.* 2009). It is generally observed that photosynthesis is enhanced in response to CO<sub>2</sub> elevation in the short-term but the stimulation declines with time due to growth limitations by environmental factors such as nutrient and soil temperature (Sage 1994; Poorter 1998; Oren *et al.* 2001; Poorter and Pérez-Soba 2001; Rogers and Ellsworth 2002; Zhang and Dang 2005).

The response of leaf-level photosynthesis to [CO<sub>2</sub>] elevation reflects a combination of adjustments in biochemical capacity and changes in leaf morphology. Morphological changes in response to [CO<sub>2</sub>] elevation may involve increased carbohydrate storage, leaf thickness, and mesophyll cell number per unit leaf area (Vu *et al.* 1989; Lou *et al.* 1994). These changes may increase in the short-term but the magnitude of the increase declines over time as a result of interactions with other environmental factors (Saxe *et al.* 1998). The stimulation of plant growth by CO<sub>2</sub> enrichment varies among plant species which likely causes differences in species distribution and patterns of forest succession with

increasing atmospheric [CO<sub>2</sub>] (Zhang and Dang 2007). The morphological traits of plant responses to CO<sub>2</sub> enrichment are also influenced by other environmental factors such as soil temperature and nutrient availability (Pettersson *et al.* 1994; Keith *et al.* 1997; Zhang and Dang 2007; Cao *et al.* 2008).

The boreal forest zone is characterized by low air and soil temperatures and a short growing season. Soil temperature is a crucial factor in determining the growth rate of plants. Low soil temperatures can decrease root growth (Domisch *et al.* 2001), thereby reducing the surface area of roots for water and nutrient absorption (Aphalo *et al.* 2006). This affects biomass accumulation due to physiological drought and nutrition stress (Zhang and Dang 2007). The activities of soil microbial organisms that decompose organic matter to release nutrients are also affected by soil temperature (Lambers *et al.* 2008). However, the response of shoots and roots to low soil temperature differs among plant species. For example, Lopushinsky and Kaufmann (1984) reported that low soil temperature reduced shoot growth but completely stopped root growth in Douglas-fir. Other studies reported increases in shoot/root ratio with increasing soil temperature from 5 to over 25 °C (Larigauderie *et al.* 1991; Landhausser *et al.* 1996).

In the present study, we have investigated the interactive effects of phosphorus (P) supply and soil temperatures under ambient and elevated [CO<sub>2</sub>] on some morphological traits of white birch seedlings (*Betula papyrifera* Marsh). Growth in the boreal forest is limited by low nutrient availability, limiting biomass production and carbon uptake in the ecosystem (Tamm 1991; Strömngren and Linder 2002). Low nutrient availability also reduces the specific leaf area (SLA), resulting in decreased leaf area available for light interception and photosynthetic carbon assimilation and consequently reduced growth

rates (Lambers *et al.* 2008). P is one of the essential macronutrients required for the normal growth and development of higher plants (Lin *et al.* 2009). Although total P is abundant in many soils, its availability in the soil solution is commonly low due to its binding to soil mineral surfaces and fixation into organic forms (Kochian *et al.* 2004). Hence, P is often present in deficient quantities (Vance *et al.* 2003), and is one of the most limiting mineral nutrients to plant growth in almost all soils (Kochian *et al.* 2004). P deficiency reduces shoot growth in plants (Whiteaker *et al.* 1976). The decrease in shoot growth in P deficient plants is the result of reduction in leaf expansion and leaf initiation (Lynch *et al.* 1991; Nielsen *et al.* 2001).

As the global climate changes in response to increasing atmospheric [CO<sub>2</sub>], changes in soil temperature and nutrient availability will be inevitable and such changes will affect plants growth and distribution in the ecosystem. However, there is a lack of information on the interactive effects of soil P supply and soil temperature under current and elevated [CO<sub>2</sub>] on the morphological traits of boreal trees. Most past studies on the effect of soil temperature and nutrients on the morphological traits of trees under current and elevated [CO<sub>2</sub>] focused on nitrogen (Cao *et al.* 2008; Lou *et al.* 1994; Ambebe *et al.* 2009). This study on the response of the morphological traits of white birch to the interaction between soil temperature and P supply under the current and doubled [CO<sub>2</sub>] will provide insights for better understanding the response of the boreal trees to rising atmospheric [CO<sub>2</sub>] and its associated effects.

## MATERIALS AND METHODS

### Plant materials

White birch seeds were germinated in the Lakehead University greenhouse (Thunder Bay, Ontario, Canada). The seeds were sown in horticultural trays filled with a mixture of peat moss and vermiculite (2:1 by volume). Seedlings of uniform height were transplanted to PVC containers (31.5 cm deep, 11 cm top diameter and 9.5 cm bottom diameter) after 4 weeks of germination and moved to treatment greenhouses as described below.

### Experimental design

The experiment comprised of two CO<sub>2</sub> concentrations, ambient (360 μmol mol<sup>-1</sup>) and elevated ([720 μmol mol<sup>-1</sup>]), three phosphorus P-supply regimes (241, 493 and 951 mg/L) and three soil temperatures (7, 17 and 27° C) in a split-split-split design. CO<sub>2</sub> was the main plot, T<sub>soil</sub> the sub-plot and P the sub-sub-plot. Nitrogen and potassium concentrations were kept at 221 and 150 mg/L, respectively, in all treatments. No other minerals were provided because they were contained in sufficient amounts in the water. The seedlings were fertilized twice a week. The soil temperatures were regulated by circulating temperature-controlled water between the containers within soil temperature control boxes. The boxes were insulated so that the soil temperature was independent of the air temperature in the greenhouses. A detailed description of the system is given by Cheng *et al.* (2000). The elevation of CO<sub>2</sub> was achieved using Argus CO<sub>2</sub> generators (Argus, Vancouver, BC, Canada). The day/night temperatures were 20 – 26/ 15 – 18°C and a day length was 16-h. The natural sunlight was supplemented by using high-pressure



sodium lamps on cloudy days, early mornings, and late evenings. The minimum illumination produced was about  $660 \mu\text{mol m}^{-2} \text{s}^{-1}$ .

## **Measurements**

Seedling height and root collar diameter (RCD) were measured after 2 and 4 months of the experiment. Six seedlings per treatment combination were randomly selected for the height and RCD at each measurement time. The leaf size of randomly selected leaves was measured with a WinFolia (Regent Instrument Inc., Quebec, Canada) and the dry mass taken after oven-drying them for 48hrs at  $70^{\circ}\text{C}$  to determine the specific leaf area (SLA). The samples were then oven-dried at  $70^{\circ}\text{C}$  for 48 hrs to determine the aboveground and belowground dry biomass using an analytical balance (precision 0.001 g).

## **Data analysis**

Treatment effects were tested using analysis of variance (ANOVA) with the software Data Desk 6.0 (Data Description, Ithaca, NY). When an interaction for a parameter was significant, multiple comparisons of means were conducted using the Least Square Difference (LSD) method to identify treatment combinations that were significantly different from each other.

## RESULTS

### Growth

The interaction among  $[\text{CO}_2]$ ,  $T_{\text{soil}}$  and P had a significant on the seedling height after 2 months of treatment (Table 3.1). The high P supply generally increased the height of the seedlings in all treatments, but the effect was not statistically significant under the low  $T_{\text{soil}}$  and ambient  $[\text{CO}_2]$ . The difference in height was also not significant between the high and intermediate P under the high  $T_{\text{soil}}$  and elevated  $[\text{CO}_2]$  or between the low and high P at low or high  $T_{\text{soil}}$  and elevated  $[\text{CO}_2]$  (Fig. 3.1A). Height generally increased with increasing  $T_{\text{soil}}$ , but the difference was not statistically significant at the low and intermediate  $T_{\text{soil}}$  under ambient  $[\text{CO}_2]$  or between intermediate and high  $T_{\text{soil}}$  under elevated  $[\text{CO}_2]$  (Fig. 3.1A). The  $\text{CO}_2$  elevation significantly increased height at the low  $T_{\text{soil}}$  and all P levels, at the intermediate and high P and intermediate  $T_{\text{soil}}$ , and low P and high  $T_{\text{soil}}$  (Fig. 3.1A). The interaction among  $[\text{CO}_2]$ ,  $T_{\text{soil}}$  and P supply became insignificant after 4 months of treatment (Table 3.1). However, the interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  significantly affected the height after 4 months of treatment. The seedling height increased with increasing  $T_{\text{soil}}$  in the ambient  $[\text{CO}_2]$  while the difference in height was not significant between the intermediate and high  $T_{\text{soil}}$  under the elevated  $[\text{CO}_2]$  (Fig. 3.1B). The  $\text{CO}_2$  elevation significantly increased seedling height at the intermediate  $T_{\text{soil}}$ , but not at the low or high  $T_{\text{soil}}$ . Seedling height increased with increasing P supply after 4 months of treatment (Table 1, Fig. 3.1B).

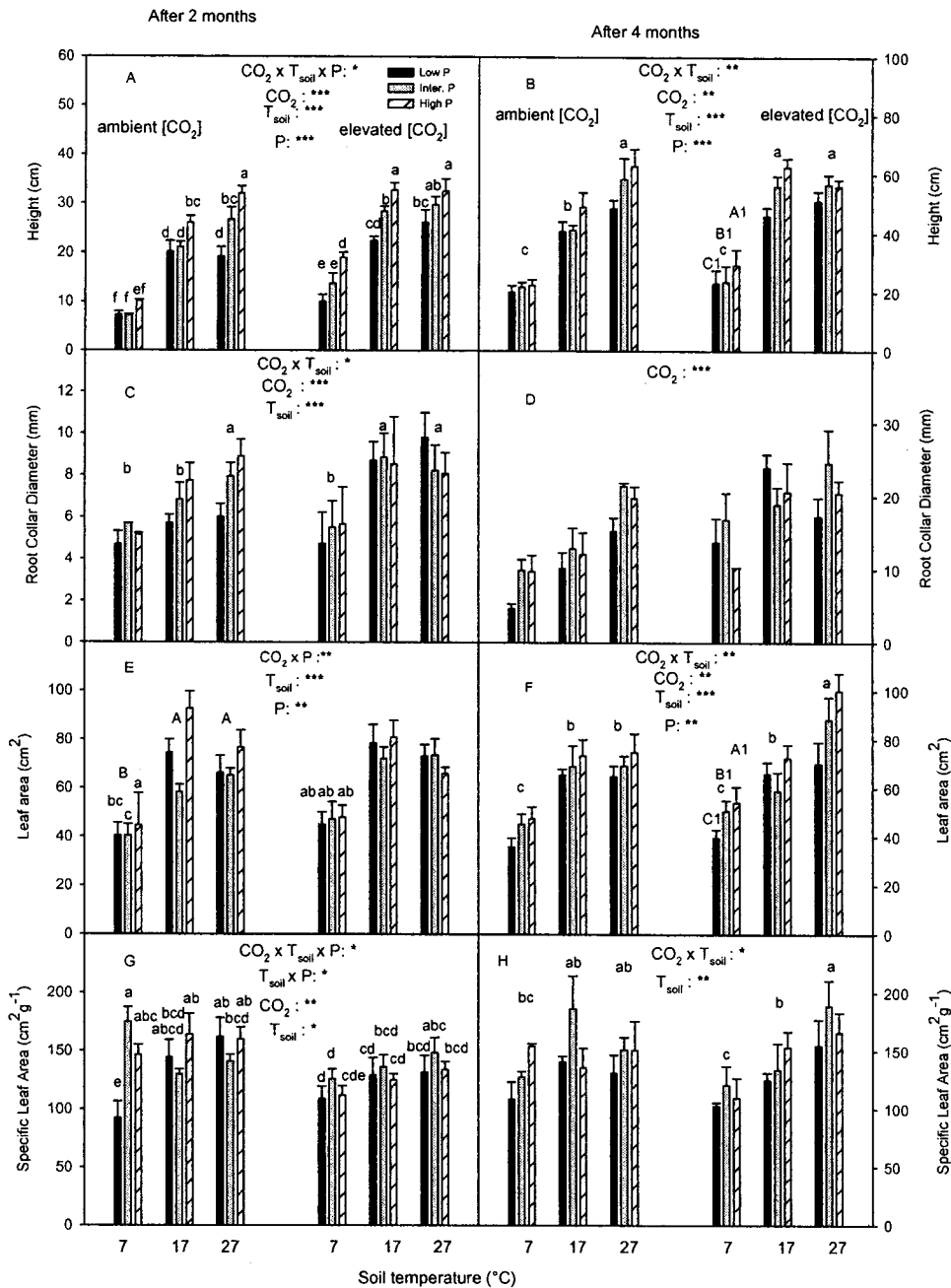
**Table 3.1.** Probabilities from ANOVA for the effects of soil temperature ( $T_{\text{soil}}$ ), phosphorus supply (P) [ $\text{CO}_2$ ] interaction on the height, RCD, leaf size and SLA in white birch seedlings. The seedlings were grown under two [ $\text{CO}_2$ ] (360 and 720  $\mu\text{mol mol}^{-1}$ ), three soil temperatures (7, 17 and 27° C) and 3 levels of P supply (241, 493 and 951 mg/L). Measurements were taken 2 and 4 months after the start of the treatment.

Source of variation	$\text{CO}_2$	$T_{\text{soil}}$	$\text{CO}_2 * T_{\text{soil}}$	P	$\text{CO}_2 * \text{P}$	$T_{\text{soil}} * \text{P}$	$\text{CO}_2 * T_{\text{soil}} * \text{P}$
After 2 months of treatment							
Height	<0.0001	<0.0001	0.4189	<0.0001	0.7085	0.5726	0.0942
RCD	0.0001	<0.0001	0.0826	0.1696	0.3199	0.4002	0.7179
Leaf area	0.2738	<0.0001	0.5683	0.0318	0.0364	0.1832	0.9798
SLA	0.0073	0.0686	0.8983	0.1625	0.2712	0.0677	0.1045
After 4 months of treatment							
Height	0.0174	<0.0001	0.0146	0.0008	0.9541	0.5283	0.4534
RCD	<0.0001	0.2261	0.8372	0.4050	0.4350	0.3446	0.1939
Leaf area	0.0420	<0.0001	0.0260	0.0021	0.5963	0.4818	0.6481
SLA	0.6068	0.0111	0.1021	0.1636	0.8547	0.8956	0.5379

Root collar diameter (RCD) was significantly affected by the interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  after 2 months of treatment (Table 3.1). Under ambient  $[\text{CO}_2]$ , the high  $T_{\text{soil}}$  significantly increased RCD while there was no significant difference between the intermediate and low  $T_{\text{soil}}$  (Fig. 3.1C). Under the elevated  $[\text{CO}_2]$ , RCD at the low  $T_{\text{soil}}$  was significantly smaller than those at the intermediate and high  $T_{\text{soil}}$  (Fig. 3.1C). The  $\text{CO}_2$  elevation significantly increased RCD only at the intermediate  $T_{\text{soil}}$ . After 4 months of treatment, however, the  $\text{CO}_2$  elevation significantly increased RCD at all  $T_{\text{soil}}$  while other factors had no significant effect on RCD (Fig. 3.1D).

Leaf size was significantly affected by the interaction between  $[\text{CO}_2]$  and P supply after 2 months of treatment (Table 3.1). The high P supply significantly increased the leaf size in the ambient  $[\text{CO}_2]$  but there was no significant difference between the low and intermediate P supply or between any P levels under elevated  $[\text{CO}_2]$  (Fig. 3.1E). The  $\text{CO}_2$  elevation increased leaf size at the intermediate P supply but did not have any significant effect at the low and high P supply (Fig. 3.1E). Additionally, the low  $T_{\text{soil}}$  significantly reduced the leaf size after 2 months of treatment (Table 3.1, Fig. 3.1E). After 4 months of treatment, the interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  had a significant effect on the leaf size (Table 3.1). The leaf size increased with increasing  $T_{\text{soil}}$  under the elevated  $[\text{CO}_2]$  (Fig. 3.1F). In the ambient  $[\text{CO}_2]$ , in contrast, there was no significant difference in leaf size between the intermediate and high  $T_{\text{soil}}$  (Fig. 3.1F).  $\text{CO}_2$  elevation increased leaf size at the high  $T_{\text{soil}}$  only. The leaf size also increased with increasing P supply after 4 months of treatment in both ambient and elevated  $[\text{CO}_2]$ .

The interaction among  $[\text{CO}_2]$ ,  $T_{\text{soil}}$  and P supply significantly affected the specific leaf area (SLA) after 2 months of treatment (Table 3.1). Under the ambient  $[\text{CO}_2]$ , the low P supply substantially decreased SLA at the low  $T_{\text{soil}}$  (Fig. 3.1G). The SLA at all P supplies and the intermediate and high  $T_{\text{soil}}$  showed no significant difference from one another. The  $\text{CO}_2$  elevation increased SLA at the low P and low  $T_{\text{soil}}$  but greatly reduced SLA at the intermediate P and low  $T_{\text{soil}}$ .  $\text{CO}_2$  elevation also reduced SLA at the high P and intermediate  $T_{\text{soil}}$  but did not affect SLA at the other treatments. The interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$  had a significant effect on SLA after 4 months of treatment (Table 3.1). The  $T_{\text{soil}}$  did not significantly affect SLA in the ambient  $[\text{CO}_2]$  while SLA increased with increasing  $T_{\text{soil}}$  under the elevated  $[\text{CO}_2]$  (Fig. 3.1H). The  $\text{CO}_2$  elevation did not significantly affect SLA at any  $T_{\text{soil}}$ .



**Fig. 3.1.** Effects of  $T_{soil}$  and P under current and doubled  $[CO_2]$  on the height, RCD, leaf size and SLA (mean + SE,  $n=6$ ) of white birch seedlings after 2 and 4 months of treatment. The seedlings were grown under two  $CO_2$  concentrations ( $360$  and  $720 \mu mol mol^{-1}$ ), three soil temperatures ( $7$ ,  $17$  and  $27^\circ C$ ) and 3 levels of P supply ( $241$ ,  $493$  and  $951 mg/L$ ). The significance levels are:  $*** = P \leq 0.001$ ,  $** = P \leq 0.05$ , and  $* = P \leq 0.10$ . Lower case letters above the bars represent interaction among  $[CO_2]$ ,  $T_{soil}$  and P supply, or interaction between  $[CO_2]$  and  $T_{soil}$  and, or interaction between  $[CO_2]$  and P. Upper case letters represent the effect of  $T_{soil}$ , while the upper case letter-number combination represents the effect of P supply. Means with the same letter(s) are not significantly different from each other or one another.

## Biomass

After 2 months of treatment, the CO<sub>2</sub> elevation increased the shoot dry biomass while the low T<sub>soil</sub> significantly reduced the shoot dry mass (Table 3.2, Fig 3.2A). The shoot dry biomass was also increased by the high P supply while there was no significant difference between the intermediate and low P supplies (Fig. 3.2A). However, the interaction between [CO<sub>2</sub>] and T<sub>soil</sub> significantly affected the shoot biomass after 4 months of treatment (Table 3.2). The shoot dry biomass was substantially decreased by the low T<sub>soil</sub> in the ambient [CO<sub>2</sub>] while the shoots dry mass increased with increases in T<sub>soil</sub> under the elevated [CO<sub>2</sub>] (Fig. 3.2B). The CO<sub>2</sub> elevation increased the shoot dry mass only at the high T<sub>soil</sub> (Fig. 3.2B). As in the 2-month measurement, high P supply significantly increased the shoot dry biomass while that of the low and intermediate P supply did not differ statistically (Table 3.2, Fig. 3.2B).

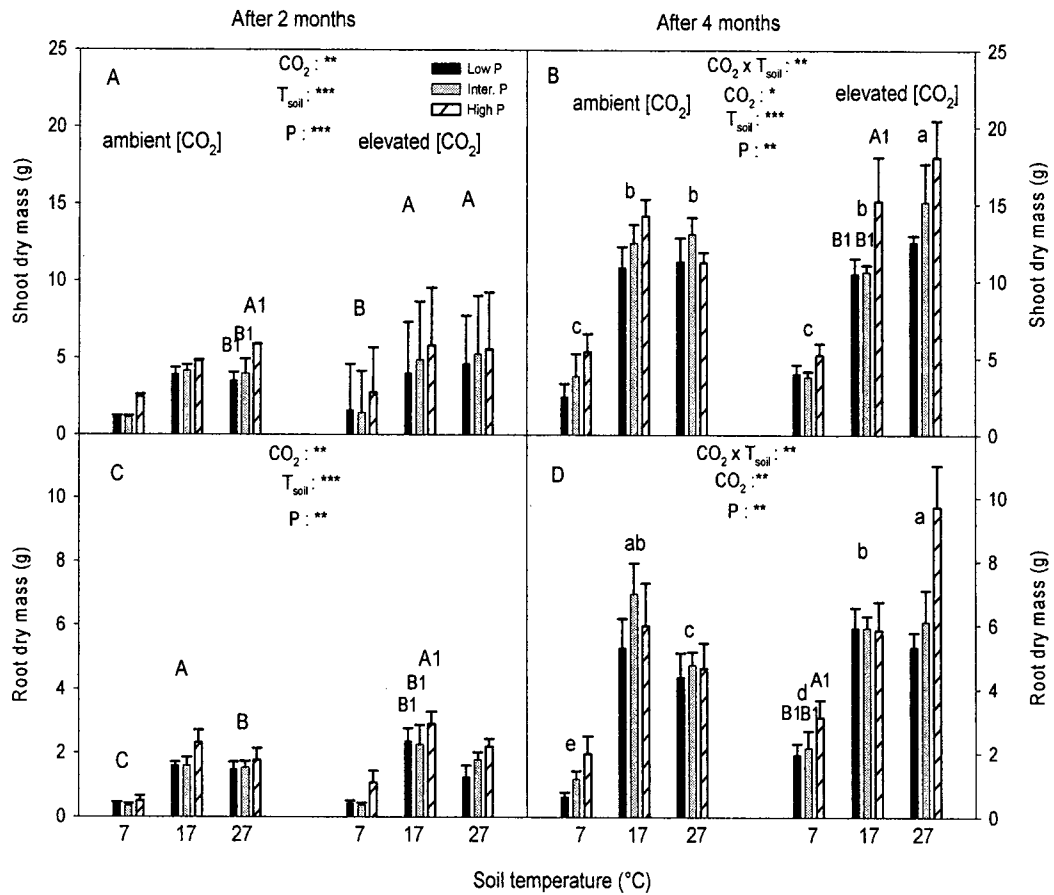
The dry root biomass was significantly affected by [CO<sub>2</sub>], T<sub>soil</sub> and P supply after 2 months of treatment (Table 3.2). The root dry biomass was lowest at low T<sub>soil</sub> and highest at the intermediate T<sub>soil</sub> (Fig. 3.2C). The high P supply significantly increased the root dry biomass but there was no significant difference between the low and intermediate P supplies. The CO<sub>2</sub> elevation significantly increased the root dry mass (Fig. 3.2C). After 4 months of treatment, the interaction between [CO<sub>2</sub>] and T<sub>soil</sub> significantly affected the root dry biomass (Table 3.2). The root dry biomass was highest at the intermediate and lowest at the low T<sub>soil</sub> under the ambient [CO<sub>2</sub>], while root mass increased with increasing T<sub>soil</sub> under the elevated [CO<sub>2</sub>] (Fig. 3.2D). The CO<sub>2</sub> elevation significantly increased the root dry biomass at the low and high T<sub>soil</sub> but did not significantly affect the dry biomass

at the intermediate  $T_{\text{soil}}$  (Fig. 3.2D). As in the measurement after 2 months, high P supply greatly increased the root dry biomass while there was no significant difference between the low and intermediate P supplies (Table 3.2, Fig. 3.2D).

**Table 3.2.** Probabilities from ANOVA for the effects of soil temperature ( $T_{\text{soil}}$ ), phosphorus supply (P) and  $[\text{CO}_2]$  interaction on shoot dry mass (SDM) and root dry mass (RDM) in white birch seedlings. Other explanations are as in Table 3.1.

Source of variation	$\text{CO}_2$	$T_{\text{soil}}$	$\text{CO}_2 * T_{\text{soil}}$	P	$\text{CO}_2 * \text{P}$	$T_{\text{soil}} * \text{P}$	$\text{CO}_2 * T_{\text{soil}} * \text{P}$
After 2 months of treatment							
SDW	0.0417	<0.0001	0.8469	0.0002	0.7947	0.8833	0.5749
RDW	0.0155	<0.0001	0.2527	0.0033	0.6053	0.8265	0.8247
After 4 months of treatment							
SDW	0.0857	<0.0001	0.0483	0.0018	0.2860	0.6577	0.4430
RDW	0.0019	<0.0001	0.0121	0.0142	0.1772	0.2357	0.1539





**Figure 3.2.** Effects of  $T_{\text{soil}}$  and P under current and doubled  $[\text{CO}_2]$  on shoot dry mass and root dry mass (mean + SE,  $n=6$ ) of white birch seedlings. The lower case letters above the bars represent the interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$ . The upper case letters above the bars represent the effect of  $T_{\text{soil}}$  and the upper case letter-number combination represents the effect of P supply. Other explanations are as in Fig. 3.1.

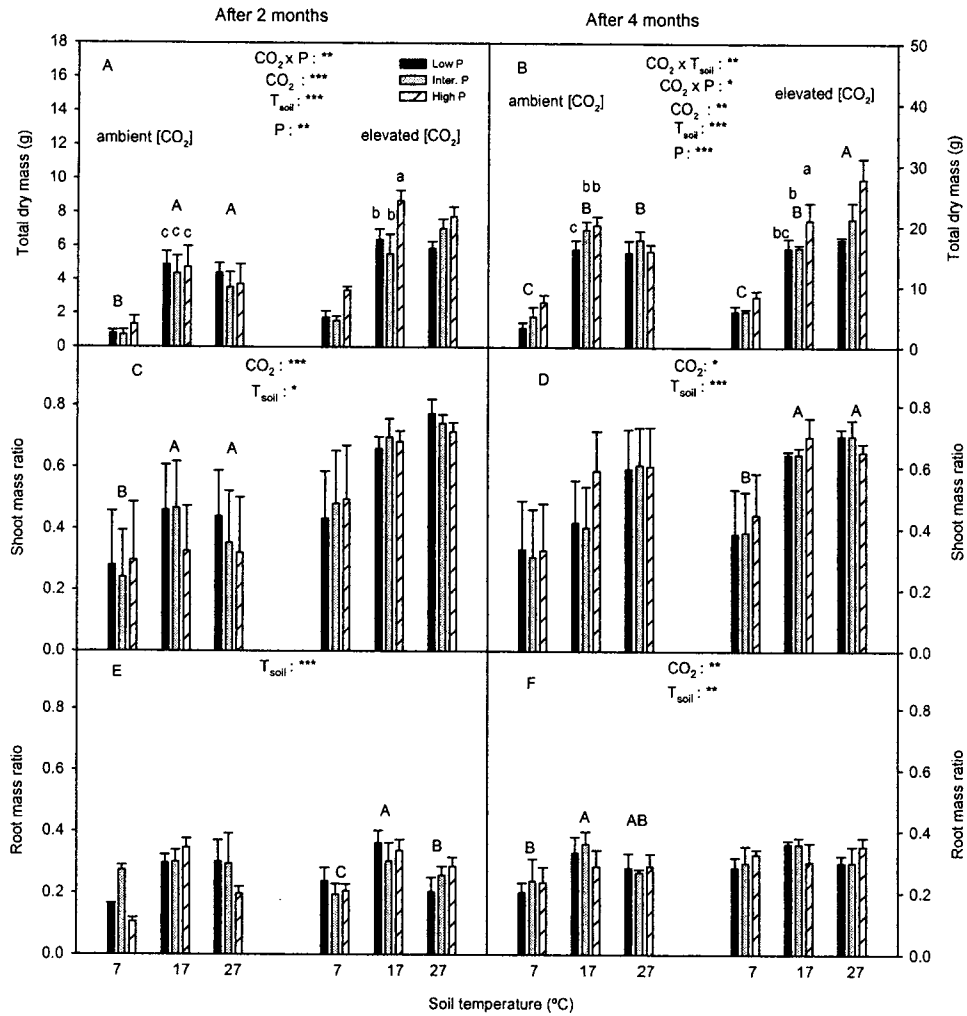
The interaction between [CO<sub>2</sub>] and P supply significantly affected the total dry biomass of the seedlings after 2 and 4 months of treatment (Table 3.3). After 2 months of treatment, there was no significant difference among the three P supplies under ambient [CO<sub>2</sub>] while the high P significantly increased the total mass under the elevated [CO<sub>2</sub>] (Fig. 3.3A). The CO<sub>2</sub> elevation increased the total dry biomass at all the P supplies, but the greatest stimulation occurred at the high P. The low T<sub>soil</sub> significantly decreased the total dry biomass after 2 months of treatment (Fig. 3.3A). After 4 months of treatment, the low T<sub>soil</sub> suppressed the biomass in both [CO<sub>2</sub>]. However, there was no significant difference between the intermediate and high T<sub>soil</sub> under the ambient [CO<sub>2</sub>]. The total dry biomass was significantly higher at the intermediate while the total mass increased with increasing T<sub>soil</sub> under elevated [CO<sub>2</sub>] (Fig 3.3B). CO<sub>2</sub> elevation significantly increased total seedling dry mass only at the high T<sub>soil</sub> (Fig. 3.3B). The interaction between [CO<sub>2</sub>] and P supply also significantly affected the total dry biomass of the seedlings after 4 months of treatment (Table 3.3). The low P supply suppressed total dry biomass under the ambient [CO<sub>2</sub>] while the total mass generally increased with P supply under the elevated [CO<sub>2</sub>], although the difference between the low and intermediate P was not statistically significant (Fig. 3.3B).

The CO<sub>2</sub> elevation significantly increased the shoot mass ratio (shoot dry biomass/total dry biomass) (SMR) after 2 months of treatment (Table 3.3, Fig 3.3C). SMR was significantly lower at the low T<sub>soil</sub> but the intermediate and high T<sub>soil</sub> did not show any significant difference after 2 months of treatment (Table 3.3, Fig 3.3C). After 4 months of treatment, the CO<sub>2</sub> elevation remained significant (Table 3.3). The lower T<sub>soil</sub> also had the lowest SMR in both ambient and elevated [CO<sub>2</sub>] (Table 3.3, Fig. 3.3D).

Root mass ratio (RMR) was significantly affected by  $T_{\text{soil}}$  after 2 months of treatment (Table 3.3). The RMR was lowest at the low  $T_{\text{soil}}$  and highest at the intermediate  $T_{\text{soil}}$  (Fig. 3.3E). After 4 months of treatment, the effect of  $T_{\text{soil}}$  remained significant (Table 3.3). RMR was highest at the intermediate  $T_{\text{soil}}$  while the low and high  $T_{\text{soil}}$  did not show any significant difference (Fig. 3.3F). The  $\text{CO}_2$  elevation, however, increased RMR after 4 months of treatment (Table 3.3, Fig. 3.3F).

**Table 3.3.** Probabilities from ANOVA for the effects of soil temperature ( $T_{soil}$ ), phosphorus supply (P) and  $[CO_2]$  interaction on total dry biomass (TDW), shoot mass ratio (SMR) and root mass ratio (RMR) in white birch seedlings. Other explanations are as in Table 3.1.

Source of variation	$CO_2$	$T_{soil}$	$CO_2 * T_{soil}$	P	$CO_2 * P$	$T_{soil} * P$	$CO_2 * T_{soil} * P$
After 2 months of treatment							
TDW	<0.0001	<0.0001	0.1226	0.0184	0.0461	0.7814	0.6547
SMR	0.0002	0.0750	0.5844	0.9214	0.8731	0.9681	0.9963
RMR	0.6510	0.0010	0.7125	0.7590	0.3319	0.6588	0.3242
After 4 months of treatment							
TWD	0.0048	< 0.0001	0.0040	0.0001	0.0989	0.9392	0.1297
SMR	0.0657	<0.0001	0.4959	0.9575	0.7251	0.8199	0.9993
RMR	0.0409	0.0347	0.4877	0.9254	0.8534	0.4288	0.9882



**Figure 3.3.** Effects of  $T_{\text{soil}}$  and P under current and doubled  $[\text{CO}_2]$  on total dry mass, shoot mass ratio (shoot dry biomass/total dry biomass) and root mass ratio (root mass/total dry biomass) of white birch seedlings (mean+ SE,  $n=6$ ). The lower case letters above the bars represent the interaction between  $[\text{CO}_2]$  and P supply. The upper case letters represent the interaction between  $[\text{CO}_2]$  and  $T_{\text{soil}}$ , interaction between  $T_{\text{soil}}$  and P supply and or the effect of  $T_{\text{soil}}$ . See Figure 3.1 for other explanations.

## DISCUSSION

Previous studies attributed the decline in the growth stimulation by CO<sub>2</sub> to photosynthetic down-regulation caused by long-term exposure to elevated [CO<sub>2</sub>] (Bazzaz *et al.* 1989; Thomas and Strain 1991). Petterson *et al.* (1993) and Rogers *et al.* (1996) reported that elevated [CO<sub>2</sub>] stimulation of growth is lesser when plants are grown with a limited nutrient supply. Our results show that the CO<sub>2</sub> elevation stimulation of height growth declined over time. At the first measurement time, the CO<sub>2</sub> elevation increased height growth at the low, intermediate and high T<sub>soil</sub> by 43, 20 12%, respectively. At the second measurement time however, height growth stimulation by elevated [CO<sub>2</sub>] especially at the low and high T<sub>soil</sub> was 16 and 3%, respectively. The decline in CO<sub>2</sub> stimulation on height growth therefore might have resulted from unfavorable low and high T<sub>soil</sub> as seedlings grew. Thus, this might have caused some stress in the seedlings as they increased in height.

There appeared to be a shift in the optimum T<sub>soil</sub> for height growth with CO<sub>2</sub> elevation. At the ambient [CO<sub>2</sub>], height growth increased with increasing but the height did not differ between the intermediate and high T<sub>soil</sub> under the elevated [CO<sub>2</sub>] indicating a lower temperature optimum under elevated [CO<sub>2</sub>]. This suggests that the pattern of plants distribution in the boreal forest might change with changes in T<sub>soil</sub> as the global atmospheric [CO<sub>2</sub>] increases.

Our results show that height growth at the low and intermediate P was generally lower than at high P and the response pattern is similar to that of other species. Lynch *et al.* (1991) and Nielsen *et al.* (2001) reported that P deficiency caused a reduction in leaf expansion and leaf initiation which resulted in reduced plant growth. Halsted and Lynch

(1996) also reported lesser growth in C<sub>3</sub> and C<sub>4</sub> plants under P stress and suggested that this might have been the result of decreased carbon fixation. We observed that seedlings at the low and intermediate P supply had smaller leaf size than those at the high P supply. This might have reduced the size of the photosynthetic machinery and the subsequent production of carbohydrates for height growth.

The CO<sub>2</sub> elevation stimulated diameter growth to a greater extent at the intermediate T<sub>soil</sub> than at the other two T<sub>soils</sub>. Interestingly, there was no significant decline of RCD growth in response to the elevated [CO<sub>2</sub>]. This suggests that more biomass was probably allocated to diameter growth than height growth. This lends some support to the prediction that growth of boreal trees will be enhanced probably due to the “fertilization” effect of CO<sub>2</sub> enrichment (Kellomaki and Wang 2001) even under unfavorable T<sub>soil</sub>. Marfo and Dang (2009) also reported an increased RCD in black spruce and white spruce under different light conditions in elevated [CO<sub>2</sub>] after four-and-a-half months of treatment. Coa *et al.* (2008) also found an increased RCD growth of 19% by CO<sub>2</sub> elevation.

The low T<sub>soil</sub> initially suppressed RCD growth in both ambient and elevated [CO<sub>2</sub>] but the effect disappeared over time. Zhang and Dang (2007) also reported a much lower diameter growth at low T<sub>soil</sub> than intermediate and high T<sub>soil</sub>. However, Lahti *et al.* (2004) reported a significantly lower diameter growth at high T<sub>soil</sub> (21°C) than at lower T<sub>soil</sub> (9°C) and suggested that carbon allocation to root growth may have been favored at the expense of shoot growth at high T<sub>soil</sub>. We found significantly lower root dry mass at the low T<sub>soil</sub> than the high T<sub>soil</sub>. We conclude that the low T<sub>soil</sub> might have suppressed growth and total biomass production.

Total seedling dry biomass was progressively increased by the intermediate and high P supply in the ambient [CO<sub>2</sub>]. At the first measurement, the percentage increase by the intermediate and high P supply was not significantly different from that of the low P. However, the percentage increases in total dry biomass by the intermediate and high P were larger and significantly different from the low P at the second measurement time. Zulu *et al.* (1991) reported that phosphorus status had large effects on plant dry mass, and total biomass was increased by 2 – to 5-fold in 6 weeks by increasing P supply. Brahim *et al.* (1996) also reported that total dry mass of 1-year-old seedlings of maritime pine (*Pinus pinaster* Ait) was dramatically decreased with P deficiency. Furthermore, the increase by the CO<sub>2</sub> elevation was also greater in the high P than in the other two P treatments.

Studies show that T<sub>soil</sub> and [CO<sub>2</sub>] have much smaller effect on biomass allocation than on growth and biomass production (Peng and Dang 2003; Zhang and Dang 2007). However, Marfo and Dang (2009) reported that CO<sub>2</sub> elevation increased shoot mass ratio (SMR) at 30% light decreased it under the 100% light condition in black spruce and white spruce. Ambebe and Dang (2009) also found that elevated [CO<sub>2</sub>] reduced biomass allocation leaf leading to lowered the leaf mass ratio in white birch seedlings. In contrast, the SMR was increased by the intermediate and high T<sub>soil</sub> and the CO<sub>2</sub> elevation, probably as a result of the increased RCD at these T<sub>soils</sub>.

It is reported that root growth is very sensitive to low T<sub>soil</sub>, which leads to reduced root extension or growth (Wan *et al.* 1999; Domisch *et al.* 2001,). High T<sub>soil</sub> also affects root physiological activities such as nutrient and water uptake and growth (Xu and Huang 2000; Huang and Fu 2001; Rachmilevitch *et al.* 2006). We observed that the intermediate



$T_{\text{soil}}$  increased RMR. The low RMR at the low and high  $T_{\text{soil}}$  might have resulted from decreased translocation of photosynthate to the root and increased accumulation of photosynthate aboveground at low  $T_{\text{soil}}$  (Lippu 1998) and/ or from high root respiratory consumption of carbohydrates at the high  $T_{\text{soil}}$  (Scheurwater *et al.* 1998; Rachmilevitch *et al.* 2006).

Our observation of increased RMR in the elevated  $[\text{CO}_2]$  is in agreement with the findings of Barrett and Gifford (1995) who observed a 20% increase in RMR in elevated  $[\text{CO}_2]$ . However, Marfo and Dang (2009) reported a lower RMR in black spruce and white spruce exposed to elevated  $[\text{CO}_2]$  and different light conditions.

The  $\text{CO}_2$  elevation mitigated the negative effect of low  $T_{\text{soil}}$  on RMR with time. We observed that the percentage reduction in RMR at the low  $T_{\text{soil}}$  in the ambient  $[\text{CO}_2]$  was 31%. However, at the elevated  $[\text{CO}_2]$ , the corresponding reduction was only 11% at the low  $T_{\text{soil}}$  after 4 months of treatment.

In conclusion, the results show that  $\text{CO}_2$  elevation partially compensated for the negative impact of low  $T_{\text{soil}}$  on height growth. There was also a shift from high to intermediate  $T_{\text{soil}}$  in the optimum  $T_{\text{soil}}$  for maximal height growth under the elevated  $[\text{CO}_2]$ . This might affect the pattern of plant distribution in the boreal forest as the atmospheric  $[\text{CO}_2]$  increases. The  $\text{CO}_2$  elevation mitigated the negative effect of low  $T_{\text{soil}}$  on biomass allocation to roots with time. With the increasing atmospheric  $[\text{CO}_2]$ , this might be beneficial to plants growing on sites where the root zone temperatures are currently too low for optimal rate of nutrients and water absorption.

## CHAPTER FOUR

### GENERAL DISCUSSION

Although  $P_n$  was higher in seedlings grown at the elevated  $\text{CO}_2$  concentrations, the extent to which seedling growth was stimulated by the  $\text{CO}_2$  elevation was proportionately less. This was because higher  $P_n$  was probably offset by lower values of SLA in the elevated  $[\text{CO}_2]$ . Previous studies with  $\text{CO}_2$  enrichment found decreases in SLA (Delucia *et al.* 1985; Pettersson and McDonald 1992; Cao *et al.* 2007). Decreased SLA at elevated  $\text{CO}_2$  might be the result of changes in leaf anatomy and/or accumulation of carbohydrates (Farrar and Williams 1991; Pettersson and McDonald 1992). In this study, lower SLA at elevated  $\text{CO}_2$  concentration was partly associated with the soil temperature and P effects on leaf growth and expansion. Other studies also attributed decline in the growth stimulation by  $\text{CO}_2$  to photosynthetic down-regulation caused by long-term exposure to elevated  $[\text{CO}_2]$  (Bazzaz *et al.* 1989; Thomas and Strain 1991) and or limited nutrient supply (Pettersson *et al.* 1993; Rogers *et al.* 1996). The decline in  $\text{CO}_2$  stimulation of height growth as observed in this study might also be attributable to increased demand for nutrients as the seedlings grew. The nutrient concentrations were not exponentially increased to match the increasing seedling growth, which may have led to lower foliar nutrient concentration.

The  $\text{CO}_2$  elevation caused a shift in the optimum  $T_{\text{soil}}$  for height growth. Height growth was significantly increased by the high  $T_{\text{soil}}$  under the ambient  $[\text{CO}_2]$  but the height did not differ between the intermediate and high  $T_{\text{soil}}$  under the elevated  $[\text{CO}_2]$ .  $\text{CO}_2$  elevation also partially mitigated the negative effect of low  $T_{\text{soil}}$  on the seedlings

height growth but the magnitude of the stimulation declined over time. We found that the seedling height growth was more resistant to high  $T_{\text{soil}}$  over time which is in agreement with the findings of Zhang and Dang (2007).

Elevated  $[\text{CO}_2]$  increased plant dry mass (Andersen *et al.* 1985; Poorter *et al.* 1996; Ishizaki *et al.* 2003; Coa *et al.* 2008). In this study, the total dry biomass production was significantly higher in the elevated  $[\text{CO}_2]$ , which supports the theory of high biomass production of plants under  $\text{CO}_2$  enrichment. However, the low  $T_{\text{soil}}$  significantly suppressed the total dry biomass production at both ambient and elevated  $[\text{CO}_2]$ . Low  $T_{\text{soil}}$  affects plant nutrient and water uptake, causing physiological nutrient stress or drought (Pastor *et al.* 1987; DeLucia *et al.* 1992; Paréz *et al.* 1993; Zhang and Dang 2007) and thereby inhibiting shoot and root growth (Folks *et al.* 1995; Peng and Dang 2003). Zhang and Dang (2007) and Ambebe and Dang (2009) also found significantly lower total biomass caused by low  $T_{\text{soil}}$  under high but not under low nutrient supply and suggested that aboveground biomass reduction was the main contributing factor.

While photosynthesis of  $\text{C}_3$  plants is generally stimulated by an increase in the atmospheric  $\text{CO}_2$  concentration, photosynthetic capacity is often reduced after long-term exposure to elevated  $\text{CO}_2$ . This reduction appears to be brought about by end-product inhibition, resulting from an imbalance in the supply and demand of carbohydrates (Arp 1991). Reduced investment in photosynthetic machinery coupled with increased carboxylation rate per unit photosynthetic machinery (carboxylation efficiency) in elevated  $[\text{CO}_2]$  may lead to down-regulation of photosynthetic capacity (Lou *et al.* 1994). In the study, it is observed that the elevated  $[\text{CO}_2]$  stimulation of  $P_n$  was higher at the

high  $T_{\text{soil}}$  but the magnitude of  $\text{CO}_2$  stimulation greatly declined with time. The measurement two months after the start of the treatment showed that  $\text{CO}_2$  elevation increased  $P_n$  by 48% but the stimulation reduced to 24% after 4 months of treatment, indicating photosynthetic down-regulation in response to the  $\text{CO}_2$  enrichment.

Decreases in leaf nutrient concentration in elevated  $\text{CO}_2$  treatments lead to a decrease in photosynthetic rate when plants are measured at the same  $\text{CO}_2$  concentration (Larigauderie *et al.* 1988). The data in this study show that the  $\text{CO}_2$  elevation reduced foliar nutrient concentration but increased their use efficiency. The lower leaf nutrient concentration at elevated  $[\text{CO}_2]$  is consistent with the theory that elevated  $[\text{CO}_2]$  can lead to the depletion of nutrient resources in plants unless they are replenished (Patterson and McDonald 1994). Decreases in nitrogen can lead to down-regulation of Rubisco activity, because it is a major component of Rubisco (Bond *et al.* 1999; Ripullone *et al.* 2003; Lewis 2004), resulting in photosynthetic down-regulation. Low phosphorus also decreases the rate of  $\text{CO}_2$  assimilation through reduction in Rubisco activity and RuBP regeneration (Brooks 1986; Jacob and Lawlor 1992; Lin *et al.* 2009). It is also involved in the transport of triose-phosphate across the chloroplast membrane and in the regulation of photophosphorylation (Flügge *et al.* 1980). Therefore, the photosynthetic down-regulation observed in the study might be attributable to the lower foliar nitrogen and phosphorus concentration in the elevated  $[\text{CO}_2]$ .

Any factor that inhibits root growth has been thought to decrease the relative stimulation of photosynthesis with increasing  $\text{CO}_2$  concentrations (Arp, 1991). However, Farrar (1988) and Ericsson *et al.* (1996) reported that growth is more sensitive to low  $T_{\text{soil}}$  than photosynthetic rates. This may be partly due to the ratio of available sinks to sources

of assimilate and the nature of feedback inhibition (Ziska 1998). In this study, it was observed that photosynthetic down-regulation of the seedlings at low  $T_{\text{soil}}$  was by 41% as compared to 54% growth suppression by the low  $T_{\text{soil}}$  relative to the intermediate  $T_{\text{soil}}$  after 4 months of the start of the experiment. This might suggest that the seedlings growth were more sensitive to the low  $T_{\text{soil}}$  than their photosynthetic rates.

In conclusion, the study revealed that higher stimulation of  $P_n$  by elevated  $[\text{CO}_2]$  might not proportionally lead to increased growth due to lower SLA. Increases in photosynthetic rates caused by  $\text{CO}_2$  enrichment also down-regulated over time, as the lower foliar nutrient concentration might have reduced the investment in Rubisco or decreased the transport of triose-phosphate across the chloroplast membrane as a result of reduced sink strength. It was also observed that seedling growth was more sensitive to low  $T_{\text{soil}}$  than to photosynthetic rate. However, the  $\text{CO}_2$  elevation appeared to cause a shift in the optimum  $T_{\text{soil}}$  (from high to low) for growth and significantly increased seedling diameter growth at all  $T_{\text{soils}}$ . The results show that in the future warmer soil temperatures combined with increased nutrient availability and other favorable environmental conditions might increase biomass production.

## REFERENCE

- Ambebe, T. F., Q. L. Dang and J. Marfo. 2009. Low soil temperature reduces the positive effects of high nutrient supply on the growth and biomass of white birch (*Betula papyrifera* Marsh.) seedlings in ambient and elevated carbon dioxide concentrations. Botany (In press).
- Andersen, I. H., C. Dons, S. Nilsen and M. K. Haugstad. 1985. Growth, photosynthesis and photorespiration of *Lemna gibba*: responses to variation in CO<sub>2</sub> and O<sub>2</sub> concentration and photon flux density. Photosynth. Res. **6**, 85 – 96.
- Aphalo, P. J., M. Lahti, T. Lehto, T. Repo, A. Rummukainen, H. Mannerkoski and L. Finér. 2006. Responses of silver birch saplings to low soil temperature. Silva Fennica **40**(3), 429–442.
- Aphalo, P. J., M. Lahti, T. Lehto, T. Repo, A. Rummukainen, H. Mannerkoski and L. Finér. 2006. Responses of silver birch saplings to low soil temperature. Silva Fennica **40**(3), 429–442.
- Arp, W. J. (1991). Effects of source-sink relations on photosynthetic acclimation to elevated CO<sub>2</sub>. Plant, Cell and Environment **14**, 869- 875.
- Barrett, D. J. and R. M. Gifford. 1995. Photosynthetic acclimation to elevated CO<sub>2</sub> in relation to biomass allocation in cotton. J. of Biogeography **22**, 331 – 339.
- Bazzaz, F. A. 1990. The response of natural ecosystems to the rising global CO<sub>2</sub> levels. Annal Review of Ecology and Systematic **21**, 167 – 196.
- Bazzaz, F. A., K. Garbutt, E. G. Reekie, and W. E. Williams. 1989. Using growth analysis to interpret competition between C<sub>3</sub> and C<sub>4</sub> annual under ambient and elevated CO<sub>2</sub>. Oecologia **79**, 223 – 235.
- Bergh, J. and S. Linder. 1999. Effects of soil warming during spring on photosynthetic recovery in boreal Norway spruce stands. Global Change Boil. **5**, 245 – 253.
- Blackman, P. G., W. J. Davies. (1985). Root to shoot communication in maize plants and the effects of soil drying. J. Exp. Bot. **36**, 39- 48.
- Bonan, B. B. and H. H. Shugart. 1989. Environmental factors and ecological processes in boreal forests. Annu. Rev. Ecol. Syst. **20**, 1–28.
- Bond, B. J., B. T. Farnsworth, R. A. Coulombe and W. E. Winner. 1999. Foliage physiology and biochemistry in response to light gradients in conifers with varying shade tolerance. Oecologia **120**, 183 – 192.
- Bowes, G. (1993) Facing the inevitable – plants and increasing atmospheric CO<sub>2</sub>. Annual Review of Plant Physiology and Plant Molecular Biology **44**, 309–332.
- Brahim, B. M., D. Loustau, J. P. Gaudilléré and E. Saur. 1996. Effect of phosphorus deficiency on photosynthesis and accumulation of starch and soluble sugars in 1-year-old seedlings of pine (*Pinus pinaster* Ait). Ann. Sci. For. **53**, 801 – 810.
- Brooks, A. 1986. Effects of phosphorus nutrition on ribulose-1,5-bisphosphate carboxylase activation, photosynthetic quantum yield and amounts of some Calvin cycle metabolites in spinach leaves. Aust. J. Plant Physiol. **12**, 221- 237.

- Bunce, J. A. 1992. Stomatal conductance, photosynthesis and respiration of temperate deciduous tree seedlings grown outdoors at an elevated concentration of carbon dioxide. *Plant, Cell and Environ.* **15**, 541 – 549.
- Bunce, J. A. 2000. Responses of stomatal conductance to light, humidity and temperature in winter wheat and barley grown at three concentrations of carbon dioxide in the field. *Global Change Biol.*, **6**, 371–382.
- Camm, E. L. and G. J. Harper. 1991. Temporal variations in cold sensitivity of roots growth in cold-stored white spruce seedlings. *Tree Physiol.* **9**, 425 – 431.
- Campbell, C. D. and R. F. Sage. 2006. Interactions between the effects of atmospheric CO<sub>2</sub> content and P nutrition on photosynthesis in white lupin (*Lupinus albus* L). *Plant, Cell and envir.* **29**, 844- 853.
- Cao, B., Q.L. Dang and S. Zhang. 2007. Relationship between photosynthesis and leaf nitrogen concentration in ambient and elevated [CO<sub>2</sub>] in white birch seedlings. *Tree physiology* **27**, 891- 899.
- Cao, B., Q.L. Dang, X. Yu and S. Zhang. 2007. Effects of [CO<sub>2</sub>] and nitrogen on morphological and biomass traits of white birch (*Betula papyrifera*) seedlings. *Forest Ecology and Management* **254**, 217 – 224.
- Coleman, J. S., F. A. Bazzaz. 1992. Effects of CO<sub>2</sub> and temperature on growth and resource use of co-occurring C<sub>3</sub> and C<sub>4</sub> annuals. *Ecology* **73**(4), 1244 - 1259.
- Convoy, J. P., P. J. Milham M. L. Reed and E. W.R. Barlow. 1990. Increase in phosphorus requirements for CO<sub>2</sub>-enriched pine species. *Plant Physiol.* **92**, 977 – 982.
- Crous, K. Y., M. B. Walters and D. S. Ellsworth. 2008. Elevated CO<sub>2</sub> concentration affects leaf photosynthesis–nitrogen relationships in *Pinus taeda* over nine years in FACE. *Tree Physiology* **28**, 607- 614.
- Davey, P. A., A. J. Parson, L. Atkinson, K. Wadge and S. P. Long. 1999. Does photosynthetic acclimation to elevated CO<sub>2</sub> increase photosynthetic nitrogen-use efficiency? A study of three native U.K. grass species in open-top chambers. *Funct. Ecol.* **13**, 21 – 28.
- Day, T. A., S. A. Heckathorn, E. H. DeLucia. 1991. Limitations of photosynthesis in *Pinus taeda* L. (Loblolly pine) at low soil temperatures. *Plant Physiol.* **96**, 1246– 1254.
- DeLucia, E. H. 1987. The effect of freezing nights on photosynthesis, stomatal conductance in seedlings of Engelmann spruce. *Plant Cell and Environ.* **10**, 333–338.
- DeLucia, E. H., S. A. Heckathorn and T. A. Day. 1992. Effects of Soil Temperature on Growth, Biomass Allocation and Resource Acquisition of *Andropogon gerardii* Vitman. *New Phytol.* **120** (4), 543- 549.
- DeLucia, E. H., T. A. Day, G. Oquist. 1991. The potential for photoinhibition of *Pinus sylvestris* L. seedlings exposed to high light and low soil temperature. *J. Exp. Bot.* **42**, 611–617.

- DeLucia, E. H., T. W. Sasek and B. R. Strain. 1985. Photosynthetic inhibition after long-term exposure to elevated levels of atmospheric CO<sub>2</sub>. *Photosynth. Res.* **7**, 175 - 184.
- Delucia, E. H., T. W. Sasek and B. R. Strain. 1985. Photosynthetic inhibition after long term exposure to elevated levels of atmospheric CO<sub>2</sub>. *Photosynthesis Research* **7**, 175 -1 84.
- Domisch, T., L. Finér and T. Lehto. 2001. Effects of soil temperature on biomass and carbohydrate allocation in Scots pine (*Pinus sylvestris*) seedlings at the beginning of the growing season. *Tree Physiology* **21**, 465–472.
- Domisch, T., L. Finér, T. Lehto and A. Smolander. 2002. Effect of soil temperature on nutrient allocation and mycorrhizas in Scots pine seedlings. *Plant and Soil* **239**, 173–185.
- Dosskey, M. G., L. Boersma and R. G. Linderma. 1993. Effect of phosphorus fertilisation on water stress in Douglas fir seedlings during soil drying. *Plant Soil* **150**, 33 – 39.
- Drake, B. G. and M. A. González-Meler. 1997. More efficient plants: a consequence of rising atmospheric CO<sub>2</sub>? *Ann. Rev. Plant. Physiol. Plant Mol. Biol.* **48**, 609 – 639.
- Eamus, D. 1991. The interaction of rising CO<sub>2</sub> and temperature with water use efficiency. *Plant, Cell and Environ.* **14**, 582 – 843.
- Eamus, D. and P. G. Jarvis. 1989. The direct effects of increase in the global atmospheric CO<sub>2</sub> concentration on natural and commercial temperate trees and forests. *Adv. Ecol. Res.* **19**, 1--55.
- Eichelmann, H., V. Oja, B. Rasulov, E. Padu, I. Bichele, H. Pettai, T. Möls, I. Kasparova, E. Vapaavuori and A. Laisk. 2004. Photosynthetic parameters of birch (*Betula pendula* Roth) leaves growing in normal and in CO<sub>2</sub>- and O<sub>3</sub>-enriched atmospheres. *Plant, Cell and Environ.* **27**, 479 – 495.
- Ellsworth, D. S. 1999. CO<sub>2</sub> enrichment in a maturing pine forest: are CO<sub>2</sub> exchange and water status in the canopy affected? *Plant, Cell and Environment* **22**, 461–472.
- Ellsworth, D. S., R. Oren, C. Huang, N. Phillips and G. R. Hendrey. 1995. Leaf and canopy responses to elevated CO<sub>2</sub> in a pine forest under Free-air CO<sub>2</sub> enrichment. *Oecologia* **104**, 139 - 146.
- Epron, D., D. Godard, G. Cornic and B. Genty. 1995. Limitation of net CO<sub>2</sub> assimilation rate by internal resistances to CO<sub>2</sub> transfer in the leaves of two tree species (*Fagus sylvatica* L. and *Castanea sativa* Mill.) *Plant Cell Environ.* **18**, 43 – 51.
- Ericsson, T., L. Rytter and E. Vapaavuori. 1996. Physiology of carbon allocation in trees. *Biom. Bioen.* **11**; 115 – 127.
- Evans, J. R. 1989. Photosynthesis and nitrogen relationships in leaves of C<sub>3</sub> plants. *Oecologia* **78**, 9 – 19.
- Evans, J. R. and F. Loreto. 2000. Acquisition and diffusion of CO<sub>2</sub> in higher plant leaves. In *Photosynthesis: Physiology and Metabolism* (Eds R. C. Leegood, T. D. Sharkey and S. von Caemmerer), pp. 321–351. Kluwer Academic Publishers, Amsterdam, The Netherlands.



- Farquhar, G.D., S. Von Caemmerer and J. A. Berry. 1980. A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. *Planta* **149**, 78-90.
- Farrar, J. F. 1988. Temperature and the partitioning and translocation of carbon. Symp. SOC Exp Biol. **42**, 203-235.
- Farrar, J. F. and M. L. Williams. 1991. The effects of increased atmospheric carbon dioxide and temperature on carbon partitioning, source-sink relations and respiration. *Plant, Cell and Environment* **14**, 819 - 830.
- Field, C. and H. A. Mooney. 1986. The photosynthesis-nitrogen relationship in wild plants. *In* On the Economy of Plant Form and Function. Ed. T. J. Givnish. Cambridge University Press, Cambridge, pp 25 – 55.
- Flügge, U. I., K. Fischer, A. Gross, W. Sebald, F. Lottspeich and C. Eckerskorn. 1989. The triose phosphate-3-phosphoglyceratephosphate translocator from spinach chloroplasts: nucleotide sequence of a full-length cDNA clone and import of the in vitro synthesized precursor protein into chloroplasts. *EMBO Journal* **8**, 39 – 46.
- Folk, R. S., S. C. Grossnickle and J. H. Russel. 1995. Gas exchange, water relations and morphology of yellow-cedar seedlings and stecklings before planting and during field establishment. *New For.* **9**, 1–20.
- Gaudillere, J. P. and M. Mousseau. 1989. Short-term effect of CO<sub>2</sub> enrichment on leaf development and gas exchange of young poplars (*Populus euramericana* cv. 1214). *Oecol. Plant.* **10**, 95—105.
- Gavito, M.E., P.S. Curtis, T.N. Mikkelsen and I. Jakobsen. 2001. Interactive effects of soil temperature, atmospheric carbon dioxide and soil N on root development, biomass and nutrient uptake of winter wheat during vegetative growth. *J. Exp. Bot.* **362**, 1913 - 1923.
- Genty, B., J. M. Briantais and N. K. Baker. (1989). The relationship between the quantum of photosynthetic electron and quenching of chlorophyll fluorescence. *Biochim. Biophys. Acta.* **990**, 87- 92.
- Gifford, R. M. 1982. Global photosynthesis in relation to our food and energy needs. *In Photosynthesis II: Development, Carbon Metabolism, and Plant Productivity* (ed, R, Govindjee), pp, 459-49, 5, Academic Press, New York.
- Goodfellow, D. 2004. Combustion method for total carbon, sulfur and nitrogen and, Nitric/Hydrochloric digestion for total phosphorus and other elements.
- Grulke, N.E., J. L. Hom and S. W. Roberts. 1993. Physiological adjustment of two full-sib families of ponderosa pine to elevated CO<sub>2</sub>. *Tree Physiol.* **12**, 391 - 401.
- Gunderson, C., R. Norby and S. Wullschlegger (2000) Acclimation of photosynthesis and respiration to simulated climatic warming in northern and southern populations of *Acer saccharum*: Laboratory and field evidence. *Tree Physiology* **20**, 87–95.
- Hällgren, J. E., M. Strand and T. Lundmark. 1991. Temperature stress. *In Physiology of Trees* (Ed. Raghavendra, AS) pp. 301 – 335. John Wiley, New York.
- Halsted, M. and J. Lynch. 1996. Phosphorus responses of C<sub>3</sub> and C<sub>4</sub> species. *J. of Experimental Botany.* **47**(297), 497- 505.

- Harper, G. J. and E. L. Camm. 1993. Effects of frozen storage duration and soil temperature on the stomatal conductance and net photosynthesis of *Picea glauca* seedlings. *Can. J. Forest Res.* **23**, 2459–2466.
- Heldt, H.W., C. J. Chon and G. H. Lorimer. 1978. Phosphate requirement for the light activation of ribulose-1.5-bisphosphate carboxylase in intact spinach chloroplast. *FEBS Letters.* **92**, 234 - 240.
- Herrick, J. D., H. Maherali and R. B. Thomas. 2004. Reduced stomatal conductance in sweet gum (*Liquidambar styraciflua*) sustained over long term CO<sub>2</sub> enrichment. *New Phytol.* **162**, 387–396.
- Huang, B. and J. Fu. 2001. Growth and physiological responses of tall fescue to surface soil drying. *International Turfgrass Society Research Journal* **9**, 291–296.
- IPCC. 2007. Summary for Policymakers. *In Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change.* Eds. S. Solomon, D. Qin, M. Manning, Z. Chen, M. Marquis, K.B Averyt, M. Tignor and H.L. Miller. Cambridge University Press, Cambridge.
- Ishizaki, S., K. Hikosaka and T. Hirose. 2003. Increased in leaf mass per unit area benefits plant growth at elevated CO<sub>2</sub> concentration. *Annals of Bot.* **91**, 905 – 914.
- Jacob, J. and D. W. Lawlor. 1991. Stomatal and mesophyll limitation of photosynthesis in phosphate deficient sunflower, maize and wheat plants. *J. Exp. Bot.* **42**, 1003 – 1011.
- Keith, H., K. L. Jacobsen and R. J. Raison. 1997. Effect of availability, temperature and moisture on soil respiration in *Eucalyptus pauciflora* forest. *Plant and Soil* **190**, 121 – 141.
- Kellomaki, S. and K.-Y. Wang. 2001. Growth and resource use of birch seedlings under elevated carbon dioxide and temperature. *Annals of Botany* **87**, 669 - 682.
- King, J. S., K. S. Pregitzer and D. R. Zak. 1999. Clonal variation in above- and below-ground responses of *Populus tremuloides* Michaux: Influence of soil warming and nutrient availability. *Plant and Soil* **217**, 119 - 130.
- Kochian, L. V., O. A. Hoekenga and M. A. Piñeros. 2004. How do crop plants tolerate acid soils? Mechanisms of aluminum tolerance and phosphorus efficiency. *Annu Rev Plant Biol.* **55**, 459-493.
- Lahti, L. M., P. J. Aphalo, L. Finér, A. Ryyppö, T. Lehto and H. Mannerkoski. 2004. Effects of soil temperature on shoot and root growth and nutrient uptake of 5-year-old Norway spruce seedlings. *Tree Physiol.* **25**, 115–122.
- Laisk, A. and A. Sumberg. 1994. Partitioning of the leaf CO<sub>2</sub> exchange into components using CO<sub>2</sub> exchange and fluorescence measurements. *Plant Physiol.* **106**, 689 – 695.
- Lambers, H., F. S. Chapin II and T. L. Pons. 2008. *Plants physiological ecology.* Springer.

- Landhaeusser, S. M., R. W. Wein, and P. Lange. 1996. Gas exchange and growth of three arctic tree-line tree species under different soil temperature and drought preconditioning regime. *Can. J. Bot.* **74** (1), 686–1693.
- Larigauderie, A., D. W. Hilbert and W. C. Oechel. 1988. Effect of CO<sub>2</sub> enrichment and nitrogen availability on resource acquisition and resource allocation in a grass, *Bromus mollis*. *Oecologia* **77**, 544 – 549.
- Lawlor, D. W. and R. A. C. Mitchell. 2000. Crop ecosystem responses to climatic change: Wheat. In: Reddy, K. R. and H. F. Hodges (eds) *Climate Change and Global Crop Productivity*. CABI Publishing, Wallingford pp 57–80.
- Lechowicz, M. J. 1984. The effects of individual variation in physiological traits on the reproductive capacity of the common cocklebur, *Xanthium strumarium* L. *Evolution* **38**, 833 - 844.
- Leegood, R. C., D. A. Walker and C. H. Foyer. 1985. Regulation of the Benson–Calvin cycle. In: Barber N, Barker R (eds) *Photosynthetic mechanisms and the environment*, Elsevier, Amsterdam, pp 189–258.
- Lewis, J. D., K. L. Griffin, R. B. Thomas and B. R. Strain. 1994. Phosphorus supply affects the photosynthetic capacity of loblolly pine grown in elevated carbon dioxide. *Tree Physiology* **14**, 1229–1244.
- Li, F., S. Kang and J. Zhang. 2004. Interactive effects of elevated CO<sub>2</sub>, nitrogen and drought on leaf area, stomatal conductance, and evapotranspiration of wheat. *Agric. Water Management* **67**, 221- 233.
- Lin, Z. H, L. S., Chen, R. B. Chen, F. Z. Zhang, H. X. Jiang and N. Tang. 2009. CO<sub>2</sub> assimilation, ribulose-1,5-bisphosphate carboxylase/oxygenase, carbohydrates and photosynthetic electron transport probed by the JIP-test, of tea leaves in response to phosphorus supply. *BMC Plant Biology* **9**, 43.
- Liozon, R., F. W. Badeck, B. Genty, S. Meyer and B. Saugier. 2000. Leaf photosynthetic characteristics of beech (*Fagus sylvatica*) saplings during three years of exposure to elevated CO<sub>2</sub> concentration. *Tree physiol.* **20**, 239 – 247.
- Lippu, J. 1998. Assimilation and allocation of carbon in scot pine seedlings during shoot elongation and as affected by soil temperature. PhD Thesis, Univ. Helsinki Dept. For. Ecol. Publication 19, 49p.
- Lloyd, J. and G. D. Farquhar. 1996. The CO<sub>2</sub> dependence of photosynthesis, plant growth responses to elevated atmospheric CO<sub>2</sub> concentrations and their interaction with soil nutrient status. 1. General principles and forest ecosystems. *Functional Ecology* **10**, 4–32.
- Long, S. P. 1991. Modification of the response of photosynthetic productivity to rising temperature by atmospheric CO<sub>2</sub> concentrations. Has its importance been underestimated? *Plant Cell Environ.* **14**, 729–39.
- Long, S. P. and B. G. Drake. 1991. Effects of the long-term elevation of CO<sub>2</sub> concentration in the field on the quantum yield of photosynthesis of the C<sub>3</sub> sedge *Scripus olneyi*. *Plant physiol.* **96**, 221 – 226.
- Long, S. P., E. A. Ainsworth, A. Rogers and D. R. Ort. 2004. Rising atmospheric carbon dioxide: plant FACE the future. *Annu. Rev. Plant Biol.* **55**, 591 – 628.

- Lopushinsky, W. and M. R. Kaufmann. 1984. Effect of cold soil on water relations and spring growth of Douglas-fir seedlings. *For. Sci.* **3**, 628 – 634.
- Loreto, F., P. C. Harley, G. Di Marco and T. D. Sharkey. 1992. Estimation of mesophyll conductance to CO<sub>2</sub> flux by three different methods. *Plant Physiol.* **98**, 1437 - 1443.
- Loustau, D., M. Ben Brahim, J. P. Gaudillière and E. Dreyer. 1999. Photosynthetic response to phosphorus nutrition in two-year-old maritime pine seedlings. *Tree Physiol.* **19**, 707 – 715.
- Luo, Y., B. Su, W. S. Currie, J. S. Dukes. 2004. Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide concentration. *Bioscience* **54**:731–739.
- Luo, Y., C. B. Field and H. A. Mooney. 1994. Predicting responses of photosynthesis and root fraction to elevated [CO<sub>2</sub>]: interaction among carbon, nitrogen, and growth. *Plant, Cell and Environment* **17**, 1195 – 1204.
- Lynch, J. P., A. Läuchli and E. Epstein. 1991. Vegetative growth of common bean in response to phosphorus nutrition. *Crop Sci.* **31**, 380 – 387.
- Marfo, J. and Q. L. Dang. 2009. Interactive effects of carbon dioxide concentration and light on the morphological and biomass characteristics of black spruce and white spruce seedlings. *Botany* **18**, 67 – 77.
- Medlyn, B. E., C. V. M. Barton, M. S. J. Broadmeadow, R. Ceulemans, P. De Angelis, M. Forstreuter, M. Freeman, S. B. Jackson, S. Kellomärk, E. Laitat, A. Rey, P. Roberts, B. D. Sigurdsson, J. Strassmeyer, K. Wang, P. S. Curtis and P. G. Jarvis. 2001. Stomatal conductance of forest species after long-term exposure to elevated CO<sub>2</sub> concentration: a synthesis. *New Phytol.* **149**, 247 – 264.
- Muhammad Iqbal Chaudhary, M. I., J. J. Adu-Gyamfi, H. Saneoka, N. T. Nguyen, R. Suwa, S. Kanai, H. A. El-Shemy, D. A. Lightfoot and K. Fujita. 2008. The effect of phosphorus deficiency on nutrient uptake, nitrogen fixation and photosynthetic rate in mashbean, mungbean and soybean. *Acta Physiol Plant* **30**, 537–544.
- Murthy, R., P. M. Dougherty, S. J. Zarnoch and H. L. Allen. 1999. Effects of carbon dioxide, fertilization, and irrigation on photosynthetic capacity of loblolly pine trees. *Tree Physiology* **16**, 537 - 546.
- Nielsen, K. L., A. Eshel and J. P. Lynch. 2001. Effect of phosphorus availability on the carbon economy of contrasting common bean (*Phaseolus vulgaris* L) genotypes. *J. Exp. Bot.* **52**(355), 329 – 339.
- Noble, R., K. F. Jensen, B. S. Ruff and K. Loats. 1992. Response of *Acer saccharum* seedlings to elevated carbon dioxide and ozone. *Ohio J. Sci.* **92**(3), 60 – 62.
- Norby, R. J. and C. M. Iversen. 2006. Nitrogen uptake, distribution, turnover, and efficiency of use in a CO<sub>2</sub>-enriched sweetgum forest. *Ecology* **87**, 5 – 14.
- Norby, R.J., E.H. DeLucia, B. Gielen et al. 2005. Forest response to elevated CO<sub>2</sub> is conserved across a broad range of productivity. *Proc. Nat'l. Acad. Sci. USA* **102**,18,052–18,056.

- Nowak, R. S., D. S. Ellsworth and S. D. Smith. 2004. Functional responses of plants to elevated atmospheric CO<sub>2</sub> – do photosynthetic and productivity data from FACE experiments support early predictions? *New Phytol.* **162**, 253 - 280.
- Oren, R., D. S. Ellsworth, K. H. Johnson, N. Phillips, B. E. Ewers, C. Maier, K. V. R. Schäfer, H. McCarthy, G. Hendrey, S. G. McNulty and G. G. Katul. 2001. Soil fertility limits carbon sequestration by forest ecosystems in a CO<sub>2</sub>-enriched atmosphere. *Nature* **411**, 469 – 471.
- Paré, D., Y. Bergeron and C. Camiré. 1993. Changes in the forest floor of Canadian southern boreal forest after disturbance. *J. Veg. Sci.* **4**, 811– 818.
- Pastor, J., R. H. Gardener, V. H. Dale, and W. M. Post. 1987. Succession changes in nitrogen availability as a potential factor contributing to spruce decline in boreal North America. *Can. J. For. Res.* **17**, 1,394 –1,400.
- Peet, M. M., S. C. Huber and D. T. Patterson. 1986. Acclimation to high CO<sub>2</sub> in monoecious cucumbers. *Plant Physiol.* **80**, 63 - 67.
- Peng, Y. Y., and Q.L. Dang. 2003. Effects of soil temperature on biomass production and allocation in seedlings of four boreal tree species. *For. Ecol. Manag.* **180**, 1– 9.
- Peterson, R. B. 1990. Effects of irradiance on the in vivo CO<sub>2</sub>:O<sub>2</sub> specificity factor in tobacco using simultaneous gas exchange and fluorescence techniques. *Plant Physiol.* **94**, 892 – 898.
- Petterson, R., A. J. S. McDonald and I. Stadenberg. 1993. Responses of small birch plants (*Betula pendula* Roth) to elevated CO<sub>2</sub> and nitrogen supply. *Plant, Cell and Env't* **16**, 1115 – 1121.
- Petterson, R. and A. J. S. McDonald. 1992. Effects of elevated carbon dioxide concentration on photosynthesis and growth of small birch plants (*Betula pendula* Roth.) at optimal nutrition. *Plant, Cell and Environment* **15**, 911-919.
- Petterson, R., A. James and S. McDonald. 1994. Effect of nitrogen supply on the acclimation of photosynthesis to elevated CO<sub>2</sub>. *Photosyn. Res.* **39**, 389 – 400.
- Poorter, H. 1998. Do slow-growing species and nutrient-stressed plants respond relatively strongly to elevated CO<sub>2</sub>? *Global Change Biol.* **4**, 693–697.
- Poorter, H. and C. Remkes. 1990. Leaf area ratio and net assimilation rate of 24 wild species differing in relative growth rate. *Oecologia* **83**, 553 - 559.
- Poorter, H. and M. Pérez-Soba. 2001. The growth response of plants to elevated CO<sub>2</sub> under non-optimal environmental conditions. *Oecologia* **129**, 1–20.
- Rachmilevitch, S., H. Lambers and B. Huang. 2006. Root respiratory characteristics associated with plant adaptation to high soil temperature for geothermal and turf-type *Agrostis* species. *J. of Experimental Bot.*, **57**(3), 623–631.
- Rao, M. and N. Terry. 1989. Leaf phosphate status, photosynthesis and carbon partitioning in sugar beet. I. Changes in growth, gas exchange and Calvin cycle enzymes. *Plant Physiol.* **90**, 814 – 819.

- Reddy, K. R., H. F. Hodges and B. A. Kimball 2000. Crop ecosystem responses to global climate change: Cotton. In: Reddy, K. R. and H. F. Hodges (eds) *Climate Change and Global Crop Productivity*. CABI Publishing, Wallingford pp 162–187.
- Rey, A. and P. G. Jarvis. 1998. Long-term photosynthetic acclimation to increased atmospheric CO<sub>2</sub> concentration in young birch (*Betula pendula*) trees. *Tree Physiol.* **18**, 441 – 450.
- Ripullone, F., G. Grassi, M. Lauteri and J. Borghette. 2003. Leaf photosynthesis-nitrogen relationships: interpretation of different patterns between *Pseudotsuga menziesii* and *Populus xeuroamericana* in a mini-stand experiment. *Tree physiol.* **23**, 137 – 144.
- Rogers, A. and D. S. Ellsworth. 2002. Photosynthetic acclimation of *Pinus taeda* (loblolly pine) to long-term growth in elevated CO<sub>2</sub> (FACE). *Plant Cell and Environ.* **25**, 851–858.
- Rogers, A., B. U. Fischer, J. Bryant, M. Frehner, H. Blum, C. A. Raines and S. P. Long. 1998. Acclimation of photosynthesis to elevated CO<sub>2</sub> under low nitrogen nutrition is affected by the capacity for assimilate utilization. Perennial ryegrass under free-air CO<sub>2</sub> enrichment. *Plant physiol.* **118**, 683 – 689.
- Rogers, A., D. J. Allen, P. A. Davey *et al.* 2004. Leaf photosynthesis and carbohydrate dynamics of soybeans grown throughout their life-cycle under Free-Air Carbon dioxide enrichment. *Plant, Cell and Environment* **27**, 449 – 458.
- Sage R. F., W. R. Pearcy and J. R. Seemann. 1987. The nitrogen use efficiency of C<sub>3</sub> and C<sub>4</sub> plants. III Leaf nitrogen effects on the activity of carboxylating enzymes in *Chenopodium album* (L.) and *Amaranthus retroflexus* (L.). *Plant Physiol.* **85**, 355 – 359.
- Sage, R. F. 1994. Acclimation of photosynthesis to increasing atmospheric CO<sub>2</sub>: the gas exchange perspective. *Photosynth. Res.* **39**, 351–368.
- Saxe, H., D. S. Ellsworth and J. Heath. 1998. Tree and forest functioning in an enriched CO<sub>2</sub> atmosphere. *New Phytologist* **139**, 395 – 436.
- Scheurwater I, C. Cornelissen, F. Dictus, R. Welschen and H. Lambers. 1998. Why do fast- and slow-growing grass species differ so little in their rate of root respiration, considering the large differences in rate of growth and ion uptake? *Plant, Cell and Environment* **21**, 995–1005.
- Sharkey, T.D. 1985. Photosynthesis in intact leaves of C<sub>3</sub> plants: physics, physiology and rate limitations. *The Botanical Review* **51**, 53-105.
- Sivak, M. N. and D. A. Walker. 1986. Photosynthesis *in vivo* can be limited by phosphate supply. *New Phytol.* **102**:499–512.
- Smith, S. E. and D. J. Read. 1997. *Mycorrhizal symbiosis*. Second edition. Academic Press. San Diego. 605p.
- Socias, F. X., H. Medrano and T. D. Sharkey. 1993. Feedback limitation of photosynthesis of *Phaseolus vulgaris* L. grown in elevated CO<sub>2</sub>. *Plant, Cell and Environment* **16**, 81- 86.

- Stitt, M. 1991. Rising CO<sub>2</sub> levels and their potential significance for carbon flow in photosynthetic cells. *Plant, Cell and Env't* **14**, 741–762.
- Strand, M. 1995. Inhibition of photosynthesis in current-year needles of unfertilized and fertilized Norway spruce (*Picea abies* (L) Karst.) during autumn and early winter. *Tree* **9**, 332 – 340.
- Strömngren, M and S. Linder. 2002. Effect of nutrition and soil warming on stemwood production in a boreal Norway spruce stands. *Global Change Biol.* **8**, 1195 – 1204.
- Tamm, C. O. 1991. Nitrogen in Terrestrial ecosystem. *Ecological Studies* **81**, pp115. Springer-Verlag, Berlin-Heidelberg.
- Teskey, R. O., T. M. Hinckley, and C. C. Grier. 1983. Effect of interruption of flow path on stomatal conductance of *Abies amabilis*. *J. Exp. Bot.* **34**, 1251-1259.
- Thomas, R. B., and B. R. Strain. 1991. Root restriction as a factor in photosynthetic acclimation of cotton seedlings grown in elevated carbon dioxide. *Plant physiol.* **96**, 627 – 634.
- Tingey, D. T., M. G. Johnson, D. L. Phillips, D. W. Johnson and J. T. Ball. 1996. Effects of elevated CO<sub>2</sub> and nitrogen on the synchrony of shoot and root growth in ponderosa pine. *Tree Physiol.* **16**, 905 - 914.
- Tissue, D. T., K. L. Griffin, R. B. Thomas and B. R. Strain. 1995. Effects of low and elevated CO<sub>2</sub> on C<sub>3</sub> and C<sub>4</sub> annuals. II. Photosynthesis and leaf biochemistry. *Oecologia* **101**, 12 – 28.
- Troeng, E. and S. Linder. 1982. Gas exchange in a 20-year-old stand of Scots pine. I. Net photosynthesis of current and one-year-old shoots within and between seasons. *Physiologia Planta.* **54**, 7 – 14.
- Vance, C. P., C. Uhde-Stone and D. L. Allan. 2003. Phosphorus acquisition and use: critical adaptations by plants for securing a nonrenewable resource. *New Phytol.* **157**, 423-447.
- Vu, J. C. V., L. H. Allen Jr. and G. Bowes. 1989. Leaf ultrastructure, carbohydrates and protein of soybeans grown under CO<sub>2</sub> enrichment. *Environmental and Experimental Botany*, **29**, 141-147.
- Walker, D. A. and S. P. Robinson. 1978. Chloroplast and cell. A contemporary view of photosynthetic carbon assimilation. *Ber. Dtsch. Bot. Ges.* **91**, 513–526.
- Wan, X., S. Landhäusser, J. J. Zwiazek and V. J. Lieffers. 1999. Root water flow and growth of aspen (*Populus tremuloides*) at low temperatures. *Tree Physiol.* **19**, 879 – 884.
- Wang, K. Y. 1996. Canopy CO<sub>2</sub> exchange of Scots pine and its seasonal variation after four-year exposure to elevated CO<sub>2</sub> and temperature. *Agricultural and Forest Meteorology* **82**, 1 - 27.
- Ward, J. K. and B. R. strain. 1999. Elevated CO<sub>2</sub> studies: past, present and future. *Tree Physiol.* **19**; 211 – 220.
- Warren, C. R. and E. Dryer. 2006. Temperature response of photosynthesis and internal conductance to CO<sub>2</sub>: results from two independent approaches. *J. Exp. Bot.* **57**(12), 3057 – 3067.

- Warren, C. R., M. A. Adams and Z. Chen. 2000. Is photosynthesis related to concentrations of nitrogen and Rubisco in leaves of Australian native plants? *Aust. J. Plant Physiol.* **27**, 407 – 416.
- Warren, C., and M. A. Adams. 2002. Phosphorus affects the growth and partitioning of nitrogen to Rubisco in *Pinus pinaster*. *Tree Physiol.* **22**, 11 – 19.
- Whiteaker, G., G. C. Gerloff, W. H. Gabelman and D. Lindgren. 1976. Intraspecific differences in growth of beans at stress levels of phosphorus. *J. American Soc. Hort. Sci.* **101**, 472 – 475.
- Xu, Q. and B. Huang. 2000. Growth and physiological responses of creeping bentgrass to changes in air and soil temperatures. *Crop Science* **40**, 1363–1368.
- Zhang, S. and Q.L. Dang. 2005. Effects of soil temperature and elevated CO<sub>2</sub> concentration on gas exchange, in vivo carboxylation and chlorophyll fluorescence in jack pine and white birch seedlings. *Tree physiology* **25**, 609- 617.
- Zhang, S. and Q.L. Dang. 2006. Effects of carbon dioxide concentration and nutrition on photosynthetic functions of white birch seedlings. *Tree physiology* **26**, 1457- 467.
- Zhang, S. and QL. Dang. 2007. Interactive effects of soil temperature and [CO<sub>2</sub>] on morphological and biomass traits in seedlings of four boreal tree species. *Forest science* **53**(3), 453 – 460.
- Ziska, L. H. 1998. The influence of root zone temperature on photosynthetic acclimation to elevated carbon dioxide concentrations. Summary. 32–33. *Ann. Bot.* **81**; 717–721.
- Zulu, J. N., F. J. Farrar and R. Whitbread. 1991. Effects of phosphate supply on the phosphorus status, dry mass and photosynthesis of wheat infected with powdery mildew. *New Phytol.* **118**, 453-461.



## APPENDIX 1

Linear Model:

$$Y_{ijkl} = \mu + C_i + \delta_{(i)} + T_j + CT_{ij} + \beta_{(ij)} + P_k + CP_{ik} + TP_{jk} + CTP_{ijk} + \varepsilon_{(ijk)l}$$

$$I = 1, 2; \quad j = 1, 2, 3; \quad k = 1, 2, 3; \quad l = 1, 2$$

Where,

$Y_{ijkl}$  = the measured response of the  $l^{\text{th}}$  replicate of the  $k^{\text{th}}$  phosphorus regime in the  $j^{\text{th}}$  soil temperature level and the  $i^{\text{th}}$  CO<sub>2</sub> concentration.

$\mu$  = the overall mean.  $C_i$  = the fixed effect of the  $i^{\text{th}}$  CO<sub>2</sub> concentration.

$\delta_{(i)}$  = the restriction error due to the restriction on the randomization of the CO<sub>2</sub> – soil temperature in the  $i^{\text{th}}$  CO<sub>2</sub> level.

$T_j$  = the fixed effect of the  $j^{\text{th}}$  soil temperature.

$CT_{ij}$  = the interaction effect of the  $j^{\text{th}}$  soil temperature in the  $i^{\text{th}}$  CO<sub>2</sub> level.

$\beta_{(ij)}$  = the restriction error due to the restriction on the randomization of the  $j^{\text{th}}$  soil temperature in the  $i^{\text{th}}$  CO<sub>2</sub> level.

$P_k$  = the fixed effect of the  $k^{\text{th}}$  phosphorus regime.

$CP_{ik}$  = the interaction effect of the  $k^{\text{th}}$  phosphorus regime in the  $i^{\text{th}}$  CO<sub>2</sub> level.

$TP_{jk}$  = the interaction effect of the  $k^{\text{th}}$  phosphorus regime in the  $j^{\text{th}}$  soil temperature.

$CTP_{ijk}$  = the interaction effect of the  $k^{\text{th}}$  phosphorus regime in the  $j^{\text{th}}$  soil temperature and the  $i^{\text{th}}$  CO<sub>2</sub> level.

$\varepsilon_{(ijk)l}$  = the random effect of the single sub-plot of the  $l^{\text{th}}$  replicate in the  $k^{\text{th}}$  phosphorus regime of the  $j^{\text{th}}$  soil temperature and the  $i^{\text{th}}$  CO<sub>2</sub> level.

**APPENDIX 1 CONT'D**

	2 3 3 2		df
	F F F R		
	i j k l	EMS	
$C_i$	0 3 3 2	$\sigma^2 + 18\sigma^2 \delta + 18\Phi C$	1
$\delta_{(i)}$	0 3 3 2	$\sigma^2 + 18\sigma^2 \delta$	0
$T_j$	2 0 3 2	$\sigma^2 + 12\Phi(T)$	2
$CT_{ij}$	0 0 3 2	$\sigma^2 + 6\sigma^2 \beta + 6 \Phi(CT)$	2
$\beta_{(ij)}$	0 0 3 2	$\sigma^2 + 6\sigma^2 \beta$	0
$P_k$	2 3 0 2	$\sigma^2 + 12\Phi(P)$	2
$CP_{ik}$	0 3 0 2	$\sigma^2 + 6\Phi_{CP}$	2
$TP_{jk}$	2 0 0 2	$\sigma^2 + 4\Phi_{TP}$	4
$CTP_{ijk}$	0 0 0 2	$\sigma^2 + 2\Phi_{CTP}$	4
$\epsilon_{(ijk)l}$	1 1 1 1	$\sigma^2$	18